The Organisms and Factors Affecting Outcomes of External Ventricular Drainage Catheter-Related Ventriculitis: A Penang Experience

Abstract

Introduction: Ventriculostomy-related infection (VRI) from external ventricular drain (EVD) insertion is a common complication and carries a high mortality rate. Choice of empiric antibiotics depends on the institutions common causative organisms and their susceptibility. We determined risk factors for mortality in patients with VRI, the common organisms causing VRI, and the rate of EVD-related VRI at our institution. **Methods:** Medical records and operative data of patients with cerebrospinal fluid positive cultures with an EVD inserted from 2012 to 2015 were traced. Forty-five patients with EVD-related VRI were included in the study. **Results:** The overall rate of VRI was 6.3%, and the overall mortality rate due to VRI was 48.9%. *Acinetobacter baumannii* was the most common organism causing VRI (14 patients, 29.2%) with a mortality rate of 64.3%. Only 14.3% of *A. baumannii* are sensitive to meropenem and imipenem. We found that patients that had a decompressive craniectomy (DC) had a lower mortality rate ($P = 0.042$) and patients with a longer duration of the EVD being in place before the diagnosis of VRI had poor outcome ($P = 0.040$). Multivariate logistic regression was performed and we found that the use of steroid ($P = 0.014$), *Pseudomonas aeruginosa* infection ($P = 0.010$), multiple organism infection ($P = 0.017$), lower Glasgow Coma Scale ($P = 0.043$), and a longer duration the EVD was in place before the diagnosis of VRI ($P = 0.008$) were related with higher mortality. **Conclusion:** VRI mortality rate is high with an alarming resistance pattern seen in *Acinetobacter* VRI. EVDs should be removed as soon as feasible, and DC may be offered to patients with severe ventriculitis or meningitis.

Keywords: *Acinetobacter, decompressive craniectomy, fatal outcome, nosocomial meningitis, risk factors, ventriculitis*

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Introduction

An external ventricular drain (EVD) is a temporary system for drainage of cerebrospinal fluid (CSF) and a conduit for intracranial pressure (ICP) monitoring. Ventriculitis, however, is a common but serious complication associated with ventricular catheters. Reported rates of EVD-related ventriculitis or ventriculostomy-related infection (VRI) range from 0% to 32.2%.[1-3] Mortality is as high as 10.3%–40.8%.[4-6] A good choice of perioperative prophylactic antibiotics and empiric antibiotics for ventriculitis while waiting for the culture report and sensitivity requires a sound knowledge of the institutions common causative organisms and their susceptibility. There is an alarming increase in resistance to antibiotics among the Gram-negative organisms.[7] *Acinetobacter* has been increasingly found to be the causative organism, and the greater

fear lies in the fact that multidrug-resistant *Acinetobacter baumannii* (MRAB) infections are getting common.[5,8] Meropenem has been the drug of choice for nosocomial meningitis and ventriculitis to cover Gram-negative bacilli, including *A. baumannii*. However, polymyxin is frequently the only available therapeutic option with the emergence of carbapenem-resistant *A. baumannii*. Since the rates of EVD-related ventriculitis can be high, and mortality is not trivial, modifiable factors affecting the outcome of these patients negatively need to be recognized. In this article, we will determine the risk factors for poor prognosis of VRI, outline the common organisms causing VRI with special attention to *Acinetobacter* VRI, and the rate of EVD-related VRI at our institution.

Methods

This was a retrospective observational study conducted at Penang General Hospital, a

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neurosurgical referral center for the Northwestern region of Peninsular Malaysia. We obtained approval from the Medical Research and Ethics Committee of Malaysia. CSF positive cultures from the neurosurgical wards were traced from the hospital's microbiology records. Hospital records and operative data of patients with CSF-positive cultures with an EVD inserted from 2012 to 2015 were traced. The total number of EVDs inserted was obtained from the neurosurgical department's records. A total of 45 patients with VRI were included in the study. Patients who had previous central nervous system (CNS) infections were excluded from the study. All patients received prophylactic ceftriaxone or cefuroxime antibiotics before EVD insertion. Most patients suspected to have ventriculitis were started on empiric antibiotics while waiting for the antibiotic susceptibility report. At our center, CSF sampling was done only when there was suspected VRI or before converting an EVD to a ventricular-peritoneal shunt.

There is no clear consensus on the definition of ventriculitis and retrospective analysis of the patient is difficult and maybe biased.[9] Therefore, ventriculitis was defined as follows: (1) any patient with a positive CSF culture from the EVD and was diagnosed and treated for ventriculitis by the neurosurgical or infectious diseases team; (2) any patient with a positive CSF culture from the EVD and increased white cells, elevated protein, and decreased CSF glucose from CSF analysis.

The Glasgow Coma Scale (GCS) is taken to be the last GCS documented before endotracheal intubation or operation. Bacteremia is defined as positive blood culture from the patients yielding the same organism as that in the CSF culture. Patients that still required the EVD even after ventriculitis was diagnosed is defined as a persistent EVD state. Date of the positive CSF culture is used to define the date when ventriculitis is confirmed. Duration of EVD when ventriculitis was first confirmed is the time in days from the date of insertion till the date of confirmation with positive CSF culture. We defined discordant empirical antibiotics as initial antibiotic choice where the cultured organism is resistant to the empiric antibiotic. We categorized the organisms into multi-resistant bacteria if they were resistant to carbapenems or were resistant to methicillin. If a single CSF sample grew more than one organism, the patient is categorized as having multiple organism growths rather than mono-organism growth.

For our statistical analyses, IBM SPSS v23.0.0 (IBM Corp) was used. Chi-square or Fisher's exact test was used for categorical variables and independent Student's *t*-test for normally distributed continuous variables, and Mann–Whitney test for nonnormally distributed continuous variables. Multivariate logistic regression analysis was also used to identify the risk factors for mortality. A statistically significant result is defined as a test showing a $P < 0.05$.

Results

The age ranged from 12 to 75 years (mean, 50.62 years). The main two underlying diagnoses leading to insertion of EVD were spontaneous intracranial bleed 75.6% and traumatic intracranial bleed 15.6%. Slightly more than half of the patients had intraventricular hemorrhage (55.6%). Decompressive craniectomy (DC) was performed on 17 (37.8%) of the patients. Only three patients were on steroids, and 12 (26.7%) patients had diabetes mellitus. Bacteremia with the same organism as that cultured in the CSF was seen in 10 (22.2%) of the patients. Out of the 45 patients, 32 (71.1%) of them still required the EVD even when ventriculitis was diagnosed. In our center, 34 (75.6%) of the patients suffered Gram-negative ventriculitis. Only three (6.7%) patients had endured recurrent infection. Discordant empirical antibiotics were given to 35 (77.8%) patients. The mortality rate from VRI for this study was 48.9%. The time from EVD insertion till confirmation of ventriculitis ranged from 2 to 21 days with a mean of 8.73 days. The mean GCS score before operation or endotracheal intubation was 7.93 with a range of 3–15 [Table 1].

During the 4-year study period of 2012–2015, a total of 796 EVDs were inserted, but 77 of them had preexisting CNS infections. The overall rate over the 4 years was 6.3% [Table 2]. This is in keeping with the published rates of 0%–32.2%.[1-3] There is a slow rise in the rate of VRI in our center, and future studies need to be taken to identify the root causes before it escalates further.

Three patients had recurrent infection, and so the total number of mono-organisms cultured was 48. *A. baumannii* and *Pseudomonas aeruginosa* together make up 50% of the causative organisms. Seven (14.6%) patients contracted coagulase negative *Staphylococcus* (CONS), and only 3 (6.2%) patients were infected with *Staphylococcus aureus* [Table 3]. Most of the *Acinetobacter* are resistant to carbapenems and only sensitive to polymyxin [Table 4]. Only 14.3% of *Acinetobacter* are sensitive to meropenem and imipenem. Ceftriaxone sensitivity is 0%, and resistance is 64.3%. As a whole, a higher proportion of *A. baumannii* is resistant with very few being sensitive to the common antibiotics. Out of the 14 patients with *A. baumannii* infection, 12 of them received discordant empiric antibiotics.

From our group of 45 patients, six of them died before specific antibiotics based on the organism susceptibility reports could be started [Table 5]. Five of the patients did not receive empiric antibiotics presumably due to a missed diagnosis initially, or the patient died before CSF analysis results could be acted on.

Discussion

Common nosocomial organisms causing infection after neurosurgical procedures include CONS, *S. aureus*,

Table 1: Characteristics of patients with external ventricular drain ventriculitis

Characteristics	*n* (%)
Sex	
Male	28 (62.2)
Female	17 (37.8)
Diagnosis	
Spontaneous intracranial bleed	33 (75.6)
Traumatic intracranial bleed	7 (15.6)
Brain tumor	2 (4.4)
Brain infarction	1 (2.2)
Pneumoventricle	1 (2.2)
IVH	
Yes	25 (55.6)
No	20 (44.4)
Indication of EVD	
ICP monitoring	2 (4.4)
CSF drainage	43 (95.6)
Craniectomy	
Yes	17 (37.8)
No	28 (62.2)
Steroid use	
Yes	3 (6.7)
No	42 (93.3)
Diabetes mellitus	
Yes	12 (26.7)
No	33 (73.3)
Bacteremia	
Yes	10 (22.2)
No	35 (77.8)
Persistent EVD	
Yes	32 (71.1)
No	13 (28.9)
CSF gram stain	
Positive	10 (22.2)
Negative	34 (75.6)
Both	1 (2.2)
Recurrent infection	
Yes	3 (6.7)
No	42 (93.3)
Discordant antibiotics	
Yes	35 (77.8)
No	10 (22.2)
Multiple organisms	
Yes	5 (11.1)
No	40 (88.9)
Multi-resistant organisms	
Yes	21 (46.7)
No	24 (53.3)
Outcome	
Alive	23 (51.1)
Dead	22 (48.9)
Total	45 (100)

EVD – External ventricular drain; CSF – Cerebrospinal fluid; IVH – Intraventricular hemorrhage; ICP – Intracranial pressure

Enterobacteriaceae, *P. aeruginosa,* and *A. baumannii*.[10-13] Although Gram-positive pathogens were the main organisms isolated in some studies, there is increasing Gram-negative pathogens being responsible for ventriculitis.[1-3,8,10,14,15] This is consistent with our study, which showed higher incidence of Gram-negative infections. *A. baumannii* and *P. aeruginosa* constituted almost half of the total organisms associated with ventriculitis in our cohort.

The name, *Acinetobacter*, comes from the Latin word for "motionless" because they lack cilia or flagella with which to move. The respiratory system is the most common site for *Acinetobacter* infection because of its transient pharyngeal colonization of healthy persons and a high rate of tracheotomy colonization, which is a common occurrence in neurosurgical patients. Most of the *A. baumannii* isolated in our study was resistant to all cephalosporins and carbapenems, but only sensitive to polymyxin. This observation raises major concern as the occurrence of these multi-resistant Gram-negative bacteria results in a significant reduction of therapeutic options for the treatment of these infections. MRAB is an emerging problem in VRI due to its ability to tolerate desiccation and to accumulate diverse mechanisms of resistance. Polymyxin is frequently the only available therapeutic option for MRAB but has poor CNS penetration. A recent study showed that only the combination of parenteral with intrathecal or intraventricular administration of polymyxin has the potential to achieve therapeutic concentrations and eradicate MRAB from the CNS.[16]

The empirical antibiotics used in our cohort consisted predominantly ceftriaxone, meropenem, and cefoperazone/sulbactam, which covered most of the Gram-negative bacteria but not MRAB [Table 5]. However, we do not advocate the use of polymyxin as first-line empirical therapy for EVD-related ventriculitis. This is because the indiscriminate use of polymyxin may contribute to the selection of further resistance and may also expose patients to unnecessary renal toxicity. Furthermore, polymyxin has weak activity against *P. aeruginosa* and other common bacteria. Therefore, careful selection of patients who should receive polymyxin as empirical therapy covering MRAB is essential. Previous colonization with *A. baumannii* resistant to carbapenems is a variable independently associated with the development of an infection caused by MRAB.[17] New ventriculitis occurring during an outbreak or in a previously colonized patient, and unresolved infections despite treatment with broad-spectrum antibiotics are the most compelling reasons for MRAB empirical coverage with polymyxin. It is important to note that *A. baumannii* transmission in neurosurgical patients is mainly due to interactions between health-care workers, patients, and contaminated fomites in the ward environment, equipment, and EVD. Infection

Table 2: Rate of external ventricular drain-related ventriculitis from 2012 to 2015

Year	Number of EVDs inserted without prior CNS infection	Number of EVDs inserted with prior CNS infection	Total number of EVDs inserted	Number of EVD-related ventriculitis	Rate of EVD-related ventriculitis (%)
2012	248	40	288	16	6.5
2013	187	12	199	8	4.3
2014	132	8	140	9	6.8
2015	152	17	169	12	7.9
Total	719	77	796	45	6.3

CNS – Central nervous system; EVDs – External ventricular drains

Table 3: Mono-organism cerebrospinal fluid culture results for single episode and recurrent episodes of ventriculitis

Pathogen	*n* (%)
Mono-organisms	
Acinetobacter baumannii	14 (29.2)
Pseudomonas aeruginosa	10 (20.8)
Coagulase negative *Staphylococcus*	7 (14.6)
Klebsiella pneumoniae	6 (12.5)
Staphylococcus aureus	3 (6.2)
Stenotrophomonas maltophilia	2 (4.1)
Corynebacterium spp	1 (2.1)
Corynebacterium macginleyi	1 (2.1)
Klebsiella oxytoca	1 (2.1)
Elizabethkingia meningoseptica	1 (2.1)
Enterobacter cloacae	1 (2.1)
Chryseobacterium indologenes	1 (2.1)
Total	48 (100)

control interventions, cohort isolation, improved hand hygiene compliance, enhanced cleaning, and environmental disinfection have been successful at reducing nosocomial infection rates and controlling outbreaks due to *A. baumannii*.[18] Ventriculitis caused by *Acinetobacter* carries a mortality rate of 29.6%–52.9%.[5,6,8,19,20] In our cohort, it is shocking that out of 14 patients with *Acinetobacter* meningitis, 9 (64.3%) of them died.

We believe our findings will create awareness among other hospitals about the rising trend of MRAB ventriculitis, which was also highlighted in other reports.[5,6,8,19,20] The level of discordance in empiric antibiotics at our center is very high (77.8%), but was statistically insignificant in terms of mortality with our small sample size (P = 0.170, odds ratio = 2.77). By knowing our center's organisms, it is our hope that there will be less wastage of antibiotics, prolongation of hospital stay, or even morbidity due to discordant antibiotics. Since there is a high rate of nosocomial MRAB ventriculitis, we will have to consider giving our patients polymyxin when they are not responding to a course of empirical antibiotics. Based on our experience, we strongly encourage every neurosurgical center to conduct a similar study to identify common causative organisms in their own ventriculitis cohort.

Various regimes of surgical prophylactic antibiotics have been tested, including cefepime, cephalothin, ampicillin/sulbactam, aztreonam, ceftaroline, trimethoprim-sulfamethoxazole, and cefuroxime.[21-25] Cefuroxime is the main surgical prophylactic antibiotic used at our center. Although there is evidence that antibiotic prophylaxis may reduce VRI, the data available are still of suboptimal quality.[26] It is postulated that the use of cephalosporins, especially aminothiazolyl cephalosporins such as cefuroxime and ceftriaxone are associated with promoting resistance due to their broad spectrum cover.[27] In view of these, it may be reasonable to use single-dose cefazolin for patients undergoing clean neurosurgical procedures, CSF-shunting procedures, or intrathecal pump placement to prevent the increase in number of resistant organisms at our center as recommended by the American Society of Health-System Pharmacists report in 2013.[28] A dosage of 1 g of cefazolin just before skin incision translates to a CSF concentration above the minimal inhibitory concentration level for approximately 5 h.[29]

There are four main postulated mechanisms of VRI by Mounier *et al.*: (1) during insertion, (2) during disconnection or manipulation of the EVD system, (3) colonization of the drain at the insertion site, and (4) hematogenous seeding. The study suggests that VRI is chiefly caused by pathogens colonizing the drain insertion site and hence, the higher number of CONS VRI.[30] Looking at the trend of organisms at our center, it is highly likely that Gram-negative infections did not occur during the insertion of EVDs. Colonization of the drain or hematogenous seeding may be a possibility since 22.2% of our patients had the same organism both in blood cultures and CSF cultures, but it could also be hematogenous dissemination of the VRI instead.

Our analysis showed that the duration of the EVD being in place before the diagnosis of VRI was made is statistically significant in determining the outcome of patients with VRI [Table 6]. The results indicated that there is a significant association between the duration of EVD being in place before the diagnosis of VRI was made and the outcome of patients with VRI (P = 0.040). It has been shown that EVD colonization was strongly associated with the development of VRI.[31] It is possible that colonization or VRI precedes detectable signs and symptoms, and therefore, the longer the EVD is within the ventricles, the higher the bacteria

Table 4: *Acinetobacter baumannii* antibiotic susceptibility from 14 mono-organism cerebrospinal fluid cultures

Antibiotic	Sensitive, *n* (%)	Intermediate, *n* (%)	Resistant, *n* (%)	Unknown, *n* (%)
Ampicillin/sulbactam	5 (35.7)	0	9 (64.3)	0
Amoxycillin/clavulanic acid	1 (7.1)	1 (7.1)	10 (71.5)	2 (14.3)
Trimethoprim/sulfamethoxazole	4 (28.6)	1 (7.1)	7 (50.0)	2 (14.3)
Gentamicin	6 (42.9)	0	8 (57.1)	0
Amikacin	7 (50.0)	0	7 (50.0)	0
Cefuroxime	0	0	9 (64.3)	5 (35.7)
Ceftazidime	2 (14.3)	0	11 (78.6)	1 (7.1)
Ceftriaxone	0	0	9 (64.3)	5 (35.7)
Cefepime	1 (7.1)	0	13 (92.9)	0
Ciprofloxacin	6 (42.9)	0	7 (50.0)	1 (7.1)
Imipenem	2 (14.3)	0	11 (78.6)	1 (7.1)
Meropenem	2 (14.3)	0	11 (78.6)	1 (7.1)
Doripenem	1 (7.1)	0	4 (28.6)	9 (64.3)
Cefoperazone/sulbactam	3 (21.4)	4 (28.6)	6 (42.9)	1 (7.1)
Piperacillin/tazobactam	1 (7.1)	0	12 (85.7)	1 (7.1)
Polymyxin B	13 (92.9)	0	0	1 (7.1)
Tigecycline	1 (7.1)	2 (14.3)	2 (14.3)	9 (64.3)

Table 5: Empiric antibiotics

Antibiotics	*n* (%)
Ceftriaxone	10 (22.2)
Meropenem	8 (17.8)
Cefoperazone/sulbactam	7 (15.6)
Cefuroxime	3 (6.7)
Piperacillin/tazobactam	3 (6.7)
Cefepime	2 (4.4)
Ampicillin/sulbactam	2 (4.4)
Imipenem	1 (2.2)
Vancomycin	1 (2.2)
Ciprofloxacin	1 (2.2)
Amoxycillin/clavulanate	1 (2.2)
Colistin and cefoperazone/sulbactam	1 (2.2)
No empiric antibiotics	5 (11.1)
Total	45 (100)

Table 6: Statistical analysis of univariate continuous variables in association with outcome

Characteristics	Alive		Dead		*P*
	n	Mean (SD)	*n*	Mean (SD)	
Duration of EVD (number of days)	23	7.0 (4.0)	22	10.0 (8.0)	0.040[a]
Age (years)	23	48.52 (15.655)	22	52.82 (16.171)	0.370[b]
GCS	23	8.17 (3.143)	22	7.68 (3.198)	0.605[b]

[a]Mann-Whitney test applied, presented as median (IQR); [b]Independent *t*-test applied. GCS – Glasgow Coma Scale; SD – Standard deviation; EVD – External ventricular drain; IQR – Interquartile range

burden is and hence more difficult to cure. On top of that, prolonged indwelling catheters may promote the selection of resistant strains. Although a persistent EVD state was not associated with a higher mortality rate in our study, it was significant in another study.[5] Yet, another paper recommends the removal of infected EVDs to improve cure rate.[32] All these information supports the early removal of EVDs if possible even if clinical VRI is not present yet. Not only does prolonged duration of EVD increase the risk of contracting VRI but it also increases the mortality rate if VRI does occur.[33]

Those who underwent a DC in our cohort of patients had a lower proportion of mortality (*P* = 0.042) [Table 7]. The mortality rate among those that had a DC was 29.4% as compared to 60.7% in those that did not have a DC. Our findings support the anecdotal evidence that DC may be beneficial in patients with bacterial meningitis with high refractory ICP.[34,35] It must be clarified that all patients in this study had the DC done for other reasons such as intracranial bleeds and not for VRI and therefore the DC was done before VRI was diagnosed. Although just a postulation, brain edema causing raised, ICP may be a cause of reduced cerebral perfusion and thus reduced antibiotic delivery and impaired transport of immune cells and factors to combat the infection. This is further supported by the fact that ICP-targeted treatment has also been recommended for bacterial meningitis.[36] In neurosurgical patients with reduced GCS and ventriculitis, it would be ideal for these patients to have their ICP monitored so that DC may be carried out if other treatment measures fail to reduce the ICP.

Multivariate logistic regression was performed, and we found that the use of steroid, *P. aeruginosa* infection, multiple organism infection, lower GCS, and a longer duration the EVD was in place before the diagnosis of VRI were related with higher mortality [Table 8]. There are various other factors that have been identified to be risk factors for poor outcome in ventricultis. In one study, mortality was strongly related to age, white cell counts, and removal of EVD.[32] Another study showed that shock, C-reactive

Table 7: Statistical analysis of univariate categorical variables in association with outcome

Characteristics	Outcome, *n* (%)		*P*	OR	95% CI	
	Alive	Dead			Lower limit	Upper limit
Craniectomy						
Yes	12 (70.6)	5 (29.4)	0.042[a]	0.27	0.07	0.98
No	11 (39.3)	17 (60.7)				
Gender						
Male	15 (53.6)	13 (46.4)	0.672[a]	0.77	0.23	2.58
Female	8 (47.1)	9 (52.9)				
IVH						
Yes	11 (44.0)	14 (56.0)	0.286[a]	1.91	0.58	6.30
No	12 (60.0)	8 (40.0)				
Indication of EVD						
ICP monitoring	1 (50.0)	1 (50.0)	0.974[b]	1.05	0.06	17.85
CSF drainage	22 (51.2)	21 (48.8)				
Steroid						
Yes	1 (33.3)	2 (66.7)	0.521[b]	2.20	0.19	26.16
No	22 (52.4)	20 (47.6)				
Diabetes						
Yes	8 (66.7)	4 (33.3)	0.208[a]	0.42	0.11	1.66
No	15 (45.5)	18 (54.5)				
Bacteremia						
Yes	5 (50.0)	5 (50.0)	0.936[b]	1.06	0.26	4.32
No	18 (51.4)	17 (48.6)				
Persistent EVD						
Yes	15 (46.9)	17 (53.1)	0.372[a]	1.81	0.49	6.76
No	8 (61.5)	5 (38.5)				
Recurrent infection						
Yes	1 (33.3)	2 (66.7)	0.521[b]	2.20	0.19	26.16
No	22 (52.4)	20 (47.6)				
Discordant antibiotics						
Yes	16 (45.7)	19 (54.3)	0.170[b]	2.77	0.61	12.51
No	7 (70.0)	3 (30.0)				
Multi-resistant organism						
Yes	8 (38.1)	13 (61.9)	0.102[a]	2.71	0.81	9.06
No	15 (62.5)	9 (37.5)				
Acinetobacter						
Yes	5 (35.7)	9 (64.3)	0.165[a]	2.49	0.68	9.19
No	18 (58.1)	13 (41.9)				
Staphylococcus aureus						
Yes	3 (75.0)	1 (25.0)	0.306[b]	0.32	0.03	3.31
No	20 (48.8)	21 (51.2)				
Coagulase negative *Staphylococcus*						
Yes	4 (57.1)	3 (42.9)	0.728[b]	0.75	0.15	3.81
No	19 (50.0)	19 (50.0)				
Klebsiella						
Yes	5 (55.6)	4 (44.4)	0.765[b]	0.80	0.18	3.47
No	18 (50.0)	18 (50.0)				
Pseudomonas						
Yes	3 (27.3)	8 (72.7)	0.069[a]	3.81	0.86	16.94
No	20 (58.8)	14 (41.2)				
Multiple organisms						
Yes	1 (20.0)	4 (80.0)	0.129[b]	4.89	0.50	47.71
No	22 (55.0)	18 (45.0)				
Gram-positive						

Contd...

Table 7: Contd...

Characteristics	Outcome, *n* (%)		*P*	OR	95% CI	
	Alive	Dead			Lower limit	Upper limit
Yes	8 (72.7)	3 (27.3)	0.099[a]	0.30	0.07	1.31
No	15 (44.1)	19 (55.9)				
Gram-negative						
Yes	16 (45.7)	19 (54.3)	0.170[b]	2.77	0.61	12.51
No	7 (70.0)	3 (30.0)				

[a]Pearson Chi-square test applied, [b]Fisher's exact test applied. OR – Odds ratio; CI – Confidence interval; EVD – External ventricular drain; IVH – Intraventricular hemorrhage; ICP – Intracranial pressure; CSF – Cerebrospinal fluid

Table 8: Multivariate logistic regression test (using enter method)

Characteristics	*P*	OR	95% CI	
			Lower limit	Upper limit
Steroid	0.014	121.420	2.646	5572.509
Pseudomonas	0.010	16.822	1.978	143.096
Multiple organism	0.017	86.766	2.211	3404.349
GCS	0.043	0.722	0.527	0.990
Duration of EVD	0.008	1.426	1.096	1.854

OR – Odds ratio; CI – Confidence interval; GCS – Glasgow Coma Scale; EVD – External ventricular drain

protein ≥10 mg/dL, and persistent EVD state was associated with higher mortality rates.[5] Gram-negative infection, CSF glucose <30 mg/dL, CSF protein >200 mg/dL, concurrent nosocomial infection, and GCS score <10 were associated with higher mortality in another study.[6]

This study has its limitations. The sample size is small even with 4 years of data, and this is a retrospective study where there maybe information bias and missing data. The study was done in a single center, and generalization of its findings to other centers is limited. Performing a prospective study and expanding the scope to include other hospitals is a suitable aim.

Conclusion

The rate of EVD-related ventriculitis in our center remains relatively low but increasing nonetheless. Worrying susceptibility patterns, especially those of *Acinetobacter* VRI needs special attention before it is too late. EVDs should be removed as soon as feasible to avoid VRI and higher mortality rates. In patients with ventriculitis/meningitis, DC maybe considered as a life-saving measure.

Conflicts of interest

There are no conflicts of interest.

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