

Access this article online
Quick Response Code:

Website: www.jlponline.org
DOI: 10.4103/JLP.JLP_148_17

# Revisiting renal amyloidosis with clinicopathological characteristics, grading, and scoring: A single-institutional experience

Abhiram Kalle, Archana Gudipati, Sree Bhushan Raju<sup>1</sup>, Karthik Kalidindi<sup>1</sup>, Swarnalatha Guditi<sup>1</sup>, Gangadhar Taduri<sup>1</sup>, Megha S. Uppin

## Abstract:

**INTRODUCTION:** Kidney involvement is a major cause of mortality in systemic amyloidosis. Glomerulus is the most common site of deposition in renal amyloidosis, and nephrotic syndrome is the most common presentation. Distinction between AA and AL is done using immunofluorescence (IF) and immunohistochemistry (IHC). Renal biopsy helps in diagnosis and also predicting the clinical course by applying scoring and grading to the biopsy findings.

**MATERIALS AND METHODS:** The study includes all cases of biopsy-proven renal amyloidosis from January 2008 to May 2017. Light microscopic analysis; Congo red with polarization; IF; IHC for Amyloid A, kappa, and lambda; and bone marrow evaluation were done. Classification of glomerular amyloid deposition and scoring and grading are done as per the guidelines of Sen S *et al.*

**RESULTS:** There are 40 cases of biopsy-proven renal amyloidosis with 12 primary and 23 secondary cases. Mean age at presentation was 42.5 years. Edema was the most common presenting feature. Secondary amyloidosis cases were predominant. Tuberculosis was the most common secondary cause. Multiple myeloma was detected in four primary cases. Grading of renal biopsy features showed a good correlation with the class of glomerular involvement.

**CONCLUSION:** Clinical history, IF, and IHC are essential in amyloid typing. Grading helps provide a subtle guide regarding the severity of disease in the background of a wide range of morphological features and biochemical values. Typing of amyloid is also essential for choosing the appropriate treatment.

## Key words:

Grading, immunofluorescence, immunohistochemistry, renal amyloidosis, typing of amyloid

## Introduction

Amyloidosis encompasses a group of diseases characterized by tissue deposition of abnormal proteins in the form of insoluble fibrils which have characteristic appearance on light microscopy, electron microscopy, and X-ray diffraction studies.<sup>[1,2]</sup> The fibrils are heterogeneous in chemical composition; Amyloid Light chain and AA are the most common, seen in primary amyloidosis or myeloma

and chronic inflammatory conditions, respectively.<sup>[2-5]</sup> Others include Amyloid transthyretin in senile systemic amyloidosis and hereditary polyneuropathies and AB2 in long-term hemodialysis.<sup>[2-6]</sup> Based on the extent of involvement, it is classified into localized and systemic forms. Most cases of hereditary amyloidosis are associated with hereditary inflammatory diseases such as familial Mediterranean fever (FMF) and Muckle–Wells syndrome.<sup>[7]</sup> Morbidity in amyloidosis is related to the direct

Departments of Pathology and <sup>1</sup>Nephrology, Nizam's Institute of Medical Sciences, Hyderabad, Telangana, India

### Address for correspondence:

Dr. Megha S. Uppin,  
Department of  
Pathology, Nizam's  
Institute of Medical  
Sciences, Punjagutta,  
Hyderabad - 500 082,  
Telangana, India.  
E-mail: megha\_harke@  
yahoo.co.in

Submission: 24-09-2017  
Accepted: 16-01-2018

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

**How to cite this article:** Kalle A, Gudipati A, Raju SB, Kalidindi K, Guditi S, Taduri G, *et al.* Revisiting renal amyloidosis with clinicopathological characteristics, grading, and scoring: A single-institutional experience. J Lab Physicians 2018;10:226-31.

toxicity of the amyloid fibrils as well as to the extent of functional compromise of the involved organ.<sup>[8,9]</sup>

The kidney is one of the most common organs to be affected in systemic amyloidosis, which is also a principal cause of mortality other than the heart, irrespective of the underlying cause.<sup>[10]</sup> The spectrum of renal morphological changes is quite variable, with the chemical composition determining the predominant site of involvement to a certain extent.<sup>[11,12]</sup> Nevertheless, glomerulus is the most common site of initial fibril deposition. Other associated findings such as glomerular sclerosis, interstitial inflammation, fibrosis, and tubular atrophy also contribute to the morbidity significantly.<sup>[13,14]</sup>

The most common presentation of renal amyloidosis is nephrotic syndrome; however, patients can present with renal failure if the deposits are predominantly vascular or medullary. It leads to end-stage renal disease if left untreated. Hypertension is less common, and diabetes insipidus has been reported uncommonly.<sup>[15,16]</sup>

Histopathology forms the cornerstone for diagnosis using Congo red staining and visualization under a polarizing microscope where it displays apple-green birefringence.<sup>[17]</sup> Tests for categorizing into primary and secondary include demonstration of clonality of light chains using immunofluorescence (IF) or immunohistochemistry (IHC) in the former and of acute-phase reactants, usually Serum Amyloid A (SAA), in the latter.<sup>[18-20]</sup> Renal biopsy is a valuable tool for diagnosis as well as predicting the clinical course.

Scoring of renal amyloid deposits has been attempted in several studies.<sup>[21-24]</sup> However, the scoring and grading scheme proposed by Sen S *et al.* provides a better means for predicting the outcomes as well as comparing therapeutic trials.<sup>[11]</sup> They have also attempted to standardize reporting of renal amyloidosis which enables uniformity and interinstitutional comparison.

The type of renal amyloidosis also has shown geographic variations. The western world has shown the dominance of primary or light-chain forms, whereas secondary form is common in the developing countries including India.<sup>[25-28]</sup>

In this article, we have tried to study the clinical features, biopsy findings, as well as grading of amyloid on renal biopsies.

## Materials and Methods

This is a retrospective study and included all cases of biopsy-proven renal amyloidosis from January 2008 to May 2017. The clinical information was retrieved from

medical records, and the clinical and demographic features were noted with respect to age and other demographic details, clinical presentation, proteinuria, renal failure, and bone marrow biopsy details.

## Histopathological analysis

Light microscopic evaluation was done with the help of hematoxylin and eosin (H and E), periodic acid–Schiff, methenamine silver–periodic acid–Schiff, Masson’s trichrome, and Congo red with polarization. In all the cases, amyloid was visualized as amorphous eosinophilic extracellular material on H and E which was congophilic and showed apple-green birefringence under polarizer.

Direct IF was performed on fresh-frozen renal biopsy using fluorescein isothiocyanate (FITC)-conjugated antibodies against kappa and lambda light chains along with IgM, IgA, IgG, C3, and C1q in 1:30 dilution followed by incubation for 30 min. The slides were examined under Zeiss Axioscope with FITC filter.

IHC was performed using ready-to-use antibodies against AA (Dako, Germany) and kappa and lambda light chains (Dako, Germany) on a fully automated immunostainer. Goat anti-rabbit anti-mouse immunoglobulin was used as a secondary antibody which was labeled using poly-horseradish peroxidase polymer, and 3,3'-diaminobenzidine was used as the chromogen.

The immunostaining, both IF and IHC, was interpreted in every case by comparing with positive and negative controls for kappa, lambda, and SAA. Tissues of documented cases were used as positive controls and tissues on which antibody addition was omitted were used as negative controls. Mesangial staining was taken into account for interpretation of light chains.

## Histopathological evaluation

The amyloid deposits in biopsies were looked for dominant involvement, i.e. glomerular, interstitial, vascular, or all compartments.

The classification of glomerular involvement from 1 to 6 along with scoring and grading is done as per the study by Sen S *et al.*

Scoring of amyloid included the extent of involvement of glomerular, interstitial, and vascular compartments as well as interstitial fibrosis and tubular atrophy, interstitial inflammation, and glomerular sclerosis in addition to the glomerular class, and the cumulative score is called the renal amyloid prognostic score (RAPS).<sup>[11]</sup> The renal findings are finally graded according to the RAPS into four grades from 0 to 3.

AL amyloidosis was confirmed by demonstrating:

- Light-chain restriction using direct IF
- Bone marrow plasma cells along with immunoelectrophoresis and immunofixation.

AA amyloidosis was considered by:

- History of chronic infection
- Absence of light-chain restriction
- Positive IHC expression of AA.

## Results

A total of 40 cases of renal amyloidosis were diagnosed in the study period including 12 primary and 23 secondary cases. Five biopsies remained unclassified due to inadequate tissue for IHC and unavailability of complete clinical details. The 40 patients included 28 male and 12 females with an M: F ratio of 2.3:1. Edema was the most common presenting feature. Others included joint pains, diarrhea, and raised serum creatinine. Mean proteinuria was 3.8 g/24 h and mean creatinine 3.1 mg/dl.

A comparison of the basic clinical and laboratory parameters is provided in Table 1. Higher mean age and light-chain restriction was seen in primary amyloidosis. Two secondary amyloidosis cases also showed lambda light-chain restriction on IF. However, these two patients had a history of chronic infection and positive AA IHC on biopsy. The bone marrow examination did not reveal increased plasma cells.

Among the 12 cases of primary amyloidosis, four were subsequently diagnosed as multiple myeloma. The remaining eight cases did not fulfill the diagnostic criteria for myeloma and hence classified as nonmyeloma-associated primary amyloidosis. Light-chain restriction in these cases was found to be lambda type, with a single case showing kappa restriction.

Tuberculosis was the most common chronic infection in the secondary amyloid cases (6/23) as shown in Graph 1.

The histopathological findings are summarized in Table 2 and show predominant glomerular involvement in all the groups.

The classification of glomerular involvement is shown in Graph 2 and Figure 1.

Grading of amyloidosis showed predominantly Grade 2 and correlated well with the class [Table 3] of glomerular involvement.

## Discussion

We report 40 cases of biopsy-proven renal amyloidosis

**Table 1: Comparison of basic clinical and biochemical parameters between primary and secondary amyloidosis**

	Primary (n=12)	Secondary (n=23)
Age (mean in years)	54	35.8
Gender (male: female)	5:1	1.5:1
Clinical features (percentage of cases)	Edema (58.33%)	Edema (52.17%)
Nephrotic proteinuria		
Number of cases	6/12	13/23
Mean (g/day)	4.16	5.46
Subnephrotic proteinuria		
Number of cases	6/12	10/23
Mean (g/day)	1.88	1.93
Normal range creatinine		
Number of cases	3/12	10/23
Mean (mg/dl)	0.83	0.73
Increased creatinine		
Number of cases	6/12	11/23
Mean (mg/dl)	3.8	4.45
Immunofluorescence pattern	Lambda: 11/12 (92%) Kappa: 1/12 (8%)	Negative: 21/23 False positive: 2/23 (lambda)

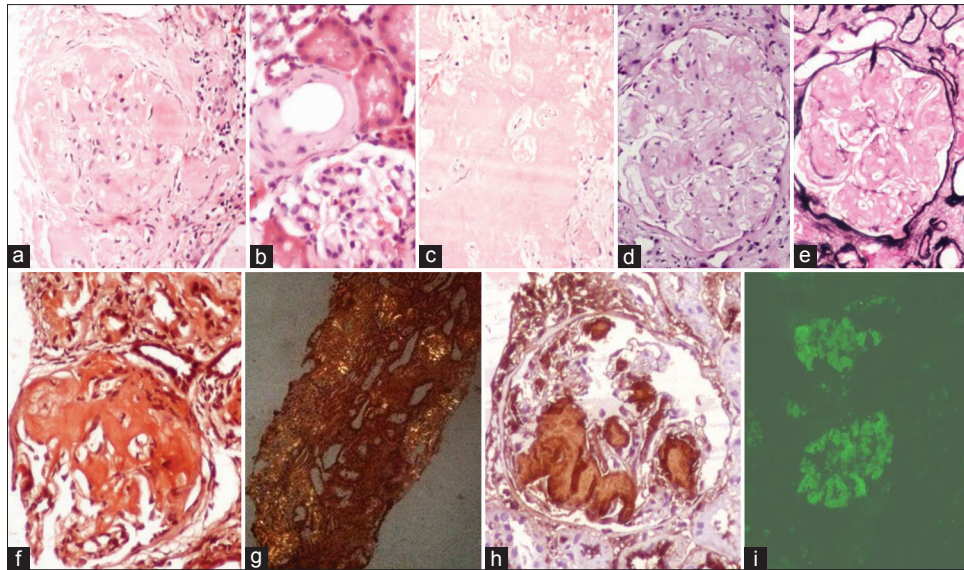
**Table 2: Light microscopic findings with respect to distribution of amyloid show glomerular deposits in the majority, both in primary as well as secondary amyloidosis groups**

Pattern of involvement	Primary (n=12) (%)	Secondary (n=23) (%)
Only glomerular	5/12 (42)	8/23 (35)
Only vascular	0	0
Only interstitial	0	0
Glomerular and vascular	6/12 (50)	12/23 (52.1)
Predominant glomerular	3	Predominant glomerular 2
Predominant vascular	1	Predominant vascular 3
Glomerular and interstitial	0	1/23 (4.3)
All compartments	1/12 (8)	2/23 (8.6)

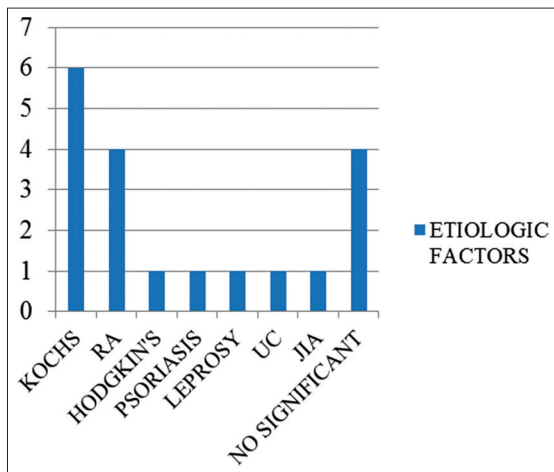
**Table 3: Grading of biopsy findings in renal amyloidosis shows majority of cases in Grade 2 and a good correlation with class of glomerular amyloid deposits**

Renal amyloid	Grade 1, number of cases (%)	Grade 2, number of cases (%)	Grade 3, number of cases (%)
Class I (n=4)	4/4 (100)	0	0
Class II (n=15)	11/15 (73)	4/15 (27)	0
Class III (n=9)	3/9 (33.3)	6/9 (66.7)	0
Class IV (n=9)	0	9/9 (100)	0
Class V (n=0)	0	0	0
Class VI (n=3)	0	2/3 (66.7)	1/3 (33.3)

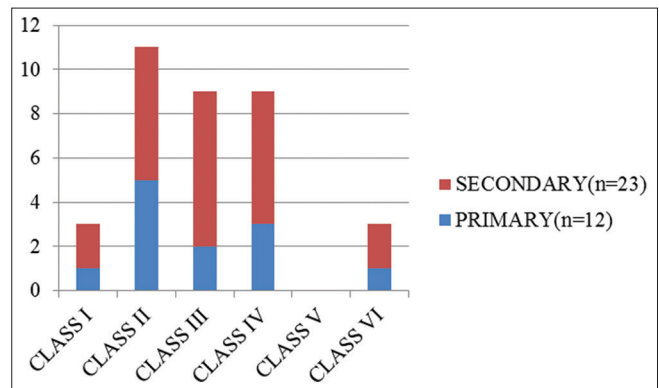




**Figure 1:** Glomerular (a), vascular (arrow) (b), and interstitial (c) amyloid deposits in H and E. These deposits are negative on periodic acid–Schiff (d) and methenamine silver-periodic acid-Schiff (e). Congo red staining shows positivity (f) with apple-green birefringence under polarizer (g). Immunohistochemistry with AA shows positivity in a case of secondary amyloidosis (h). Direct immunofluorescence showing lambda positivity in a primary amyloidosis case (i)



**Graph 1:** Spectrum of secondary causes of amyloidosis. RA=Rheumatoid arthritis; UC=Ulcerative colitis; JIA= Juvenile idiopathic arthritis



**Graph 2:** Classification of glomerular amyloid deposits shows cases predominantly in Class 2

over a 9-year period, which accounted for 0.34% of all the renal biopsies in the study period. Majority (65.7%) of the cases were secondary in our series, similar to other Indian and some of the Asian studies. Contrarily, primary amyloidosis is more common in studies from the Western world.<sup>[25-27]</sup> The mean age at presentation is variable, ranging from 35 to 63 years in different studies similar to that observed in the present study.<sup>[7,25-30]</sup> Primary amyloidosis is seen in the elderly age group. Chronic infections such as tuberculosis and leprosy are prevalent in India and other developing Asian countries, and this is perhaps the reason for more number of secondary amyloid cases. Renal involvement is the rule in secondary AA type of amyloidosis which is seen in younger individuals as against the primary amyloidosis.

Majority of our patients had nephrotic range proteinuria with subnephrotic range in few. This finding is also comparable to other studies and can be explained by the predominance of glomerular involvement.<sup>[11,12,26,31-33]</sup> A comparison of the findings of the present study with other similar studies is provided in Tables 4 and 5.

Some of the authors correlated the principal site of involvement in the renal biopsy with the type of precursor protein and showed that fibrinogen A alpha amyloidosis is typically glomerular and the deposits in AApoA and ALECT2 are extraglomerular and present with renal failure.<sup>[11,12]</sup> The pattern of renal involvement in the present study was predominantly glomerular (90%), either in isolation or in addition to other compartments. Exclusive vascular or interstitial involvement was not seen in the absence of glomerular deposition. There were no differences between primary and secondary amyloidosis cases with respect to the distribution of

**Table 4: Comparison of the basic clinical, biochemical, and histological findings of the present study with other similar Indian studies shows comparable results**

	Present study (2017) (n=40)	Usha <i>et al.</i> (2013) (n=13)	Shah <i>et al.</i> (1996) (n=75)	Chugh <i>et al.</i> (1981) (n=233)
Study period (years)	8	5	20	23
Age (mean in years)	42.5	43.5	35	35.2
Gender (male: female)	2.3:1	12:1	5.2:1	2.5:1
Clinical features (percentage of cases)	Edema (57)	Edema (84.6)	Edema (52)	Edema (72)
Chronic infection (%)	TB (26.1)	TB (27.7)	TB (79)	TB (59.1)
Nephrotic range proteinuria (mean in g/day)	4.82	7.63	-	-
Subnephrotic proteinuria (mean in g/day)	1.83	2.5	-	-
Normal range serum creatinine (mean in mg/dl)	0.92	1.22	-	-
Raised serum creatinine (mean in mg/dl)	4.66	11.3	-	-
AL:AA	1:1.9	1:2.5	1:9	1:15.6
Histology (%)	Glomerular (90)	Glomerular (84.6)	Predominantly glomerular	Predominantly glomerular

TB=Tuberculosis, AL= Amyloid light chain, AA= Amyloid A

**Table 5: Comparison of basic clinical, biochemical, and histological features of the present study with similar western studies shows a predominance of primary amyloidosis cases in the latter**

	Present study (2016) (n=35)	Sait <i>et al.</i> (2010) (n=288)	Hopfer <i>et al.</i> (2011) (n=403)	Said <i>et al.</i> (2013) (n=474)
Study period (years)	9	17	47	5
Age (mean in years)	42.5		59.5	63
Gender (male: female)	2.3:1		1.23:1	1.6:1
Clinical features (percentage of cases)	Edema (57)			Edema (78)
Nephrotic range proteinuria (mean in g/day)	4.82			6.1
Subnephrotic proteinuria (mean in g/day)	1.83			
Normal range serum creatinine (mean in mg/dl)	0.92			1.3
Raised serum creatinine (mean in mg/dl)	4.66			
AL: AA	1:1.9	1:9	1.2:1	12.3:1
Histology (%)	Glomerular (90)	Predominantly glomerular	Glomerular (84.6)	Glomerular (97)

AL= Amyloid light chain, AA= Amyloid A

amyloid in various compartments of the kidney. This is the reason why light microscopic findings alone are not sufficient to characterize the type of amyloid protein. Clinical history as well as IF and IHC is essential in the workup of amyloid subtyping.

We attempted to score and grade the amyloid as per the guidelines of Sen S *et al.*<sup>[11]</sup> Most of our cases belonged to Classes 2–4 and none belonged to Class 5. In their study, a positive correlation was noted between Class 1 and Grade 1, Class 3 and Grade 2, and Class 4 and Grade 3. Our findings were comparable to that of Sait *et al.* as has been given in Table 5. These findings help provide a subtle guide to the clinicians regarding the severity of the disease in the background of a wide range of morphological features and biochemical values. However, it needs a clinical follow-up of patients for comparison with the outcome and survival analysis. The findings of Sen S *et al.*<sup>[11]</sup> were predominantly based on AA type of precursor in cases of FMF which was the major cause of secondary amyloidosis in their study. Unlike that, the present study had a variety of underlying

conditions, with tuberculosis being the major chronic infection. Our series does not include any hereditary cases. Nevertheless, the grading of renal biopsy showed a good correlation with the class of glomerular amyloid deposits and can be expected to give a good idea of overall prognosis.

Typing of amyloid is also essential for choosing the appropriate line of treatment. The current treatment strategies aim to eradicate the underlying source of clonal plasma cell population in AL amyloidosis. Chemotherapeutic drugs and autologous stem cell transplantation to help the recovery of the marrow have been employed for the same. On the other hand, the treatment for AA amyloidosis aims to halt the chronic inflammatory process by employing cytotoxic agents and tumor necrosis factor antagonists. Even with these agents, it may be difficult to suppress SAA production adequately. Molecules that inhibit fibrillogenesis are under trial for these cases.<sup>[8]</sup> Treatment-associated toxicity with chemotherapy remains an issue of concern. Hence, therapy needs to be administered only after

the nature of amyloid is proved to be AL or AA by appropriate tests on the biopsy.

Retrospective nature of the study and absence of follow-up data are the limitations of the present study.

## Conclusion

We have seen more number of secondary amyloidosis in patients having a history of varied chronic infectious and inflammatory diseases. The pattern of renal involvement does not vary with the type of amyloidosis. Grading and scoring the amyloid on renal biopsy correlates with the class of glomerular involvement.

## Financial support and sponsorship

Nil.

## Conflicts of interest

There are no conflicts of interest.

## References

- Glenner GG. Amyloid deposits and amyloidosis. The beta-fibrilloses (first of two parts). *N Engl J Med* 1980;302:1283-92.
- Westermarck P, Benson MD, Buxbaum JN, Cohen AS, Frangione B, Ikeda S, et al. A primer of amyloid nomenclature. *Amyloid* 2007;14:179-83.
- Pepys MB. Amyloidosis. *Annu Rev Med* 2006;57:223-41.
- Sipe JD, Benson MD, Buxbaum JN, Ikeda S, Merlini G, Saraiva MJ, et al. Amyloid fibril protein nomenclature: 2012 recommendations from the Nomenclature Committee of the International Society of Amyloidosis. *Amyloid* 2012;19:167-70.
- Sipe JD, Benson MD, Buxbaum JN, Ikeda S, Merlini G, Saraiva MJ, et al. Nomenclature 2014: Amyloid fibril proteins and clinical classification of the amyloidosis. *Amyloid* 2014;21:221-4.
- Gejyo F, Yamada T, Odani S, Nakagawa Y, Arakawa M, Kunitomo T, et al. A new form of amyloid protein associated with chronic hemodialysis was identified as beta 2-microglobulin. *Biochem Biophys Res Commun* 1985;129:701-6.
- Real de Asúa D, Costa R, Contreras MM, Gutiérrez Á, Filigghedu MT, Armas M, et al. Clinical characteristics of the patients with systemic amyloidosis in 2000-2010. *Rev Clin Esp (Barc)* 2013;213:186-93.
- Dember LM. Amyloidosis-associated kidney disease. *J Am Soc Nephrol* 2006;17:3458-71.
- Keeling J, Teng J, Herrera GA. AL-amyloidosis and light-chain deposition disease light chains induce divergent phenotypic transformations of human mesangial cells. *Lab Invest* 2004;84:1322-38.
- Gertz MA. Amyloidosis: Diagnosis and prognosis. *Fut Rheumatol* 2008;3:369-80.
- Sen S, Sarsik B. A proposed histopathologic classification, scoring, and grading system for renal amyloidosis: Standardization of renal amyloid biopsy report. *Arch Pathol Lab Med* 2010;134:532-44.
- Said SM, Sethi S, Valeri AM, Leung N, Cornell LD, Fidler ME, et al. Renal amyloidosis: Origin and clinicopathologic correlations of 474 recent cases. *Clin J Am Soc Nephrol* 2013;8:1515-23.
- Sasatomi Y, Kiyoshi Y, Uesugi N, Hisano S, Takebayashi S. Prognosis of renal amyloidosis: A clinicopathological study using cluster analysis. *Nephron* 2001;87:42-9.
- Verine J, Mourad N, Desseaux K, Vanhille P, Noël LH, Beaufils H, et al. Clinical and histological characteristics of renal AA amyloidosis: A retrospective study of 68 cases with a special interest to amyloid-associated inflammatory response. *Hum Pathol* 2007;38:1798-809.
- Carone FA, Epstein FH. Nephrogenic diabetes insipidus caused by amyloid disease. Evidence in man of the role of the collecting ducts in concentrating urine. *Am J Med* 1960;29:539-44.
- Asmundsson P, Snaedal J. Persistent water diuresis in renal amyloidosis. A case report. *Scand J Urol Nephrol* 1981;15:77-9.
- Dumoulin M, Bader R. A short historical survey of developments in amyloid research. *Protein Pept Lett* 2006;13:213-7.
- Kebbel A, Röcken C. Immunohistochemical classification of amyloid in surgical pathology revisited. *Am J Surg Pathol* 2006;30:673-83.
- Strege RJ, Saeger W, Linke RP. Diagnosis and immunohistochemical classification of systemic amyloidoses. Report of 43 cases in an unselected autopsy series. *Virchows Arch* 1998;433:19-27.
- Schönland SO, Heigenbart U, Bochtler T, Mangatter A, Hansberg M, Ho AD, et al. Immunohistochemistry in the classification of systemic forms of amyloidosis: A systematic investigation of 117 patients. *Blood* 2012;119:488-93.
- Watanabe T, Saniter T. Morphological and clinical features of renal amyloidosis. *Virchows Arch A Pathol Anat Histol* 1975;366:125-35.
- Shiiki H, Shimokama T, Yoshikawa Y, Toyoshima H, Kitamoto T, Watanabe T, et al. Renal amyloidosis. Correlations between morphology, chemical types of amyloid protein and clinical features. *Virchows Arch A Pathol Anat Histopathol* 1988;412:197-204.
- Shiiki H, Shimokama T, Yoshikawa Y, Onoyama K, Morimatsu M, Watanabe T, et al. Perimembranous-type renal amyloidosis: A peculiar form of AL amyloidosis. *Nephron* 1989;53:27-32.
- Nagata M, Shimokama T, Harada A, Koyama A, Watanabe T. Glomerular crescents in renal amyloidosis: An epiphenomenon or distinct pathology? *Pathol Int* 2001;51:179-86.
- Pinney JH, Smith CJ, Taube JB, Lachmann HJ, Venner CP, Gibbs SD, et al. Systemic amyloidosis in England: An epidemiological study. *Br J Haematol* 2013;161:525-32.
- Shah VB, Phatak AM, Shah BS, Kandalkar BM, Haldankar AR, Ranganathan S, et al. Renal amyloidosis – A clinicopathologic study. *Indian J Pathol Microbiol* 1996;39:179-85.
- Gertz MA, Kyle RA. Secondary systemic amyloidosis: Response and survival in 64 patients. *Medicine (Baltimore)* 1991;70:246-56.
- Dixit R, Gupta R, Dave L, Prasad N, Sharma S. Clinical profile of patients having pulmonary tuberculosis and renal amyloidosis. *Lung India* 2009;26:41-5.
- Tuglular S, Yalcinkaya F, Paydas S, Oner A, Utas C, Bozfkioğlu S, et al. A retrospective analysis for aetiology and clinical findings of 287 secondary amyloidosis cases in Turkey. *Nephrol Dial Transplant* 2002;17:2003-5.
- Bergesio F, Ciciani AM, Santostefano M, Brugnano R, Manganaro M, Palladini G, et al. Renal involvement in systemic amyloidosis – An Italian retrospective study on epidemiological and clinical data at diagnosis. *Nephrol Dial Transplant* 2007;22:1608-18.
- Usha, Singh RG, Parkash J, Kapoor R, Rai S, Sinha DK. Clinicopathological study of renal amyloidosis. *Clinicopathological study of renal amyloidosis*. *JK Sci* 2006;8:18-23.
- Chugh KS, Datta BN, Singhal PC, Jain SK, Sakhuja V, Dash SC, et al. Pattern of renal amyloidosis in Indian patients. *Postgrad Med J* 1981;57:31-5.
- Hopfer H, Wiech T, Mihatsch MJ. Renal amyloidosis revisited: Amyloid distribution, dynamics and biochemical type. *Nephrol Dial Transplant* 2011;26:2877-84.