

Which platelet function test best reflects the *in vivo* plasma concentrations of ticagrelor and its active metabolite?

The HARMONIC study

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Summary

Aim of this study was assessment of the relationship between concentrations of ticagrelor and its active metabolite (AR-C124910XX) and results of selected platelet function tests. In a single-centre, cohort study, patients with myocardial infarction underwent blood sampling following a 180 mg ticagrelor loading dose intake (predose, 1, 2, 3, 4, 6, 12, 24 hours postdose) to perform pharmacokinetic and pharmacodynamic assessments. Platelet reactivity was evaluated using the VASP-assay, the VerifyNow device and the Multiplate analyzer. Analysis of 36 patients revealed high negative correlations between ticagrelor concentrations and platelet reactivity evaluated with all three platelet function tests (the VASP-assay: $RS=-0.722$; $p<0.0001$; the VerifyNow device: $RS=-0.715$; $p<0.0001$; the Multiplate analyzer: $RS=-0.722$; $p<0.0001$), with no significant differences between correlation coefficients. Similar results were found for AR-C124910XX. Platelet reactivity values assessed with all three methods generally correlated well with

each other; however, a significantly higher correlation ($p<0.02$) was demonstrated between the VerifyNow and Multiplate tests ($RS=0.707$; $p<0.0001$) than in other assay combinations (the VASP-assay and the VerifyNow device: $RS=0.595$; $p<0.0001$; the VASP-assay and the Multiplate analyzer: $RS=0.588$; $p<0.0001$). With respect to the recognition of high platelet reactivity, we found higher measurement concordance between the VerifyNow and Multiplate tests compared with other assay combinations, while for low platelet reactivity, only results of the VerifyNow and Multiplate assay were related to each other. Platelet reactivity measurements performed with the VASP, VerifyNow and Multiplate tests show comparably strong negative correlations with ticagrelor and AR-C124910XX concentrations.

Keywords

Ticagrelor, AR-C124910XX, flow cytometric VASP-assay, VerifyNow device, Multiplate analyzer

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Introduction

Ticagrelor is a direct-acting, reversible platelet P2Y₁₂ receptor inhibitor recommended by both European and American guidelines as first line therapy in a broad spectrum of patients with acute coronary syndromes (ACS) (1, 2). The faster, stronger and more consistent inhibition of platelet function by ticagrelor and its active metabolite (AR-C124910XX) translated into improved patient outcomes in the landmark PLATO trial (3, 4). Importantly, multiple platelet function tests are usually simultaneously used in pharmacodynamic studies on P2Y₁₂ receptor inhibitors. Additionally, platelet function testing is a tool which may aid in better clinical decision making (5). According to both European and American ex-

perts, there are three recommended platelet function tests, namely the VerifyNow device, the Multiplate analyzer and the vasodilator-stimulated phosphoprotein (VASP) phosphorylation assay, and none of these three methods is preferred over another (6–8).

Notably, due to the reversibility of the receptor binding as well as fast onset and offset of action, plasma concentrations of ticagrelor and AR-C124910XX well reflect the extent of platelet P2Y₁₂ receptor inhibition (3, 9). So far there are no studies linking measurements of ticagrelor and AR-C124910XX concentrations with comparative evaluation of methods used for platelet reactivity assessment.

The primary goal of this trial was to evaluate the relationship between plasma concentrations of ticagrelor and AR-C124910XX

on the one hand and the results of all three recommended platelet function tests on the other in patients with acute myocardial infarction (AMI). The secondary study goal included assessment of correlation and agreement between the results of the three compared platelet function tests.

Methods

Study design

The study was designed as a prospective, single-centre, investigator-initiated, cohort trial. Consecutive AMI patients admitted to The Department of Cardiology and Internal Medicine between 6:00 a.m. and 6:00 p.m. received orally a 300 mg loading dose (LD) of plain aspirin (Polpharma SA, Starogard Gdański, Poland) and were screened with respect to their eligibility for the study enrollment. Each patient provided written informed consent for participation prior to any study specific procedures. Inclusion criteria were as follows: age between 18 and 80 years, male or non-pregnant female patients with a diagnosis of ST-segment elevation AMI (STEMI) or non-ST-segment elevation AMI (NSTEMI), and

provision of informed consent for angiography and percutaneous coronary intervention (PCI). Exclusion criteria were: treatment with any P2Y₁₂ receptor inhibitor within 14 days prior to the study enrollment, ongoing treatment with oral anticoagulant or chronic therapy with low-molecular-weight heparin, active bleeding, Killip class III or IV during screening for eligibility, respiratory failure, and history of coagulation disorders (10). Additionally, we pre-specified that patients receiving a glycoprotein (GP) IIb/IIIa receptor inhibitor will be excluded from the primary analysis as this therapy may affect the results of platelet reactivity assessment with the VerifyNow and Multiplate tests. STEMI and NSTEMI were diagnosed in line with The Third Universal Definition of Myocardial Infarction (11). All study participants received orally a 180 mg LD of ticagrelor with 250 ml of tap water immediately after baseline blood sampling. Subsequently, within 15 minutes (min) from the ticagrelor LD, all patients underwent a coronary angiography assessment followed by PCI, if necessary. The study participants were given a 90 mg ticagrelor maintenance dose every 12 hours (h) and a 75 mg aspirin maintenance dose every 24 h following the LDs. The study was approved by the Local Ethics Committee in accordance with the Helsinki Declaration.

Table 1: Baseline characteristics of study participants. Data are shown as median (interquartile range) or number (%). AMI – acute myocardial infarction; CABG – coronary artery bypass grafting; n/a – not applicable; NSTEMI – non-ST-segment elevation myocardial infarction; PCI – percutaneous coronary intervention; STEMI – ST-segment elevation myocardial infarction.

Variable	Overall study population (n=45)	Patients included in the primary analysis (n=36)	Patients not included in the primary analysis (n=9)	P-value
Age [years]	62.0 (55.0–72.0)	62.0 (55.5–73.5)	59.0 (55.0–67.0)	0.477
Female	11 (24.4)	8 (22.2)	3 (33.3)	0.666
Body mass index [kg/m ²]	26.9 (24.8–30.1)	26.7 (24.6–30.2)	26.9 (26.5–28.4)	0.650
STEMI/NSTEMI	31 (68.9)/14 (31.1)	22 (61.1)/14 (38.9)	9 (100)/0 (0)	0.024
Hypertension	20 (44.4)	18 (50)	2 (22.2)	0.261
Diabetes mellitus	9 (20)	8 (22.2)	1 (11.1)	0.661
Dyslipidaemia	40 (88.9)	31 (86.1)	9 (100)	n/a
Current smoker	19 (42.2)	15 (41.7)	4 (44.4)	1.00
Prior AMI	7 (15.6)	7 (19.4)	0	n/a
Prior PCI	8 (17.8)	8 (22.2)	0	n/a
Prior CABG	0	0	0	n/a
Prior non-severe heart failure	1 (2.2)	1 (2.8)	0	n/a
Prior non-haemorrhagic stroke	0	0	0	n/a
Peripheral arterial disease	2 (4.4)	2 (5.6)	0	n/a
Chronic renal disease	2 (4.4)	2 (5.6)	0	n/a
Chronic obstructive pulmonary disease	0	0	0	n/a
Gout	3 (6.6)	2 (5.6)	1 (11.1)	0.500
Morphine use in the acute phase of AMI	21 (46.7)	15 (41.7)	6 (66.7)	0.267

Study endpoints

Blood samples were collected at multiple time points in order to assess study endpoints in a broad range of ticagrelor/AR-C124910XX concentrations and values of platelet reactivity. For each study participant, eight values of ticagrelor/AR-C124910XX concentrations and platelet reactivity, if not stated otherwise, were included in the final analysis. Values of correlation coefficients or intra-class correlation coefficients were compared between the assessed platelet function tests.

The primary study endpoint was the correlation coefficient between ticagrelor concentrations and platelet reactivity measurements, while the secondary study endpoints included:

- correlation coefficient between AR-C124910XX concentrations and platelet reactivity measurements,
- correlation coefficient between ticagrelor concentrations and inhibition of platelet reactivity (inhibition of platelet reactivity was defined as pre-LD of ticagrelor platelet reactivity minus actual platelet reactivity, divided by pre-LD of ticagrelor platelet reactivity; for each study participant, seven values of ticagrelor concentration and inhibition of platelet reactivity (for sampling points at 1, 2, 3, 4, 6, 12, 24 h post-LD) were included in the final analysis),
- correlation coefficient between AR-C124910XX concentrations and inhibition of platelet reactivity (additional description – same as above),
- Bland-Altman analysis assessing agreement between the standardised values of platelet reactivity measured with the compared platelet function tests (standardized platelet reactivity was defined as actual platelet reactivity minus mean platelet reactivity, divided by standardised deviation of platelet reactivity),
- correlation coefficient between platelet reactivity measured with the compared platelet function tests,

- intra-class correlation coefficient between platelet reactivity measured with the compared platelet function tests,
- κ statistic to assess agreement between platelet reactivity measurements with the compared platelet function tests in terms of identifying patients with high platelet reactivity (HPR) or low platelet reactivity (LPR).

Additionally, we decided to perform an exploratory, post-hoc sub-analysis comparing STEMI and NSTEMI patients.

Blood sampling

Blood samples were collected using a venous catheter (18G) inserted into a forearm vein at eight pre-defined time points (prior to the LD of ticagrelor and 1, 2, 3, 4, 6, 12 and 24 h post-LD). The first 3–5 ml of blood were discarded to avoid spontaneous platelet activation.

Measurement of ticagrelor and AR-C124910XX concentrations

Ticagrelor and AR-C124910XX plasma concentrations were measured separately using liquid chromatography coupled with tandem mass spectrometry as described in our previous publication (12). Lower limits of quantification were 4.69 ng/ml both for ticagrelor and AR-C124910XX.

Assessment of platelet reactivity

Platelet reactivity was evaluated using the flow cytometric VASP-assay (VASP; BioCytex, Marseille, France), the VerifyNow device (VerifyNow P2Y12 assay, Accumetrics, Inc., San Diego, CA, USA) and multiple electrode aggregometry (MEA) with the Multiplate analyzer (ADPtest, Roche Diagnostics International Ltd.,

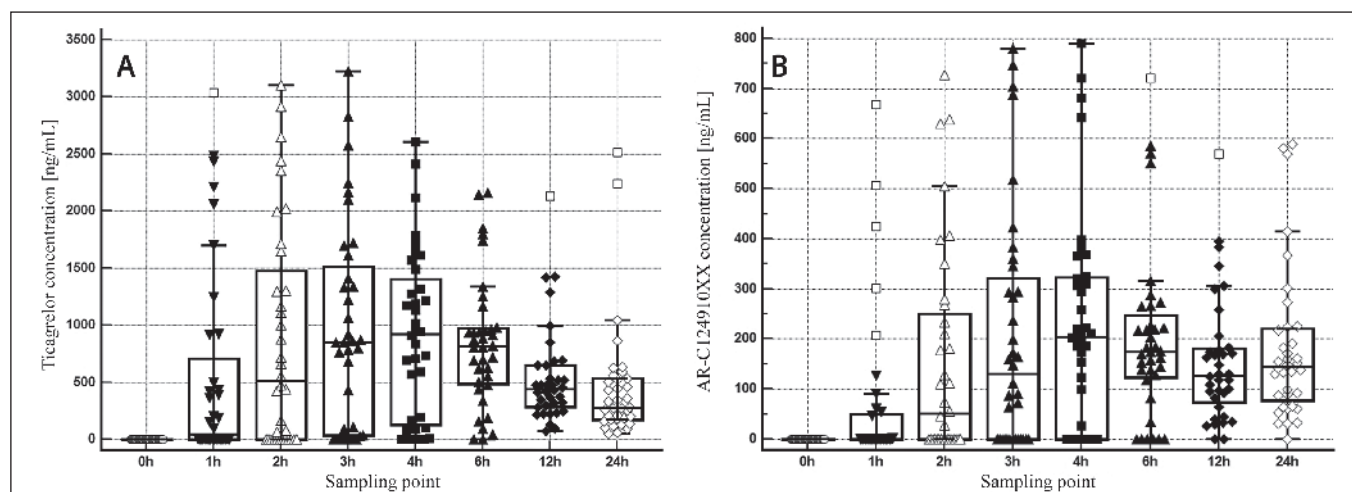
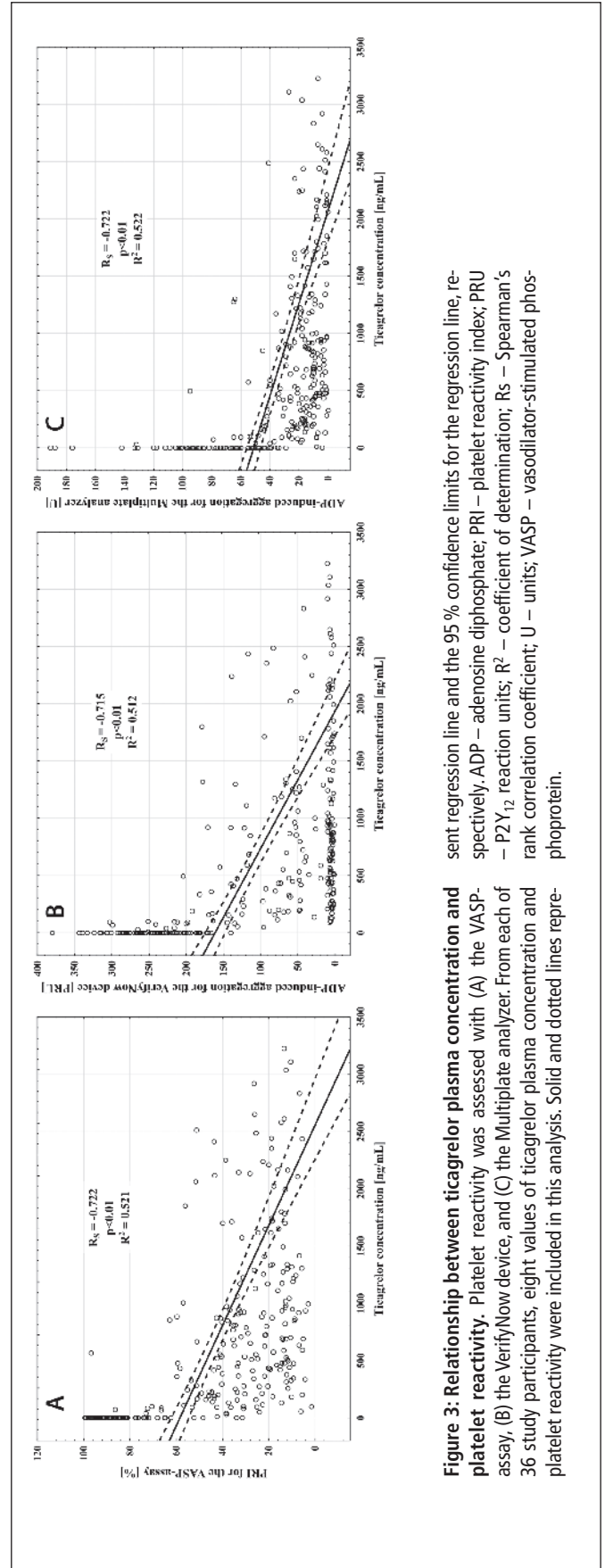
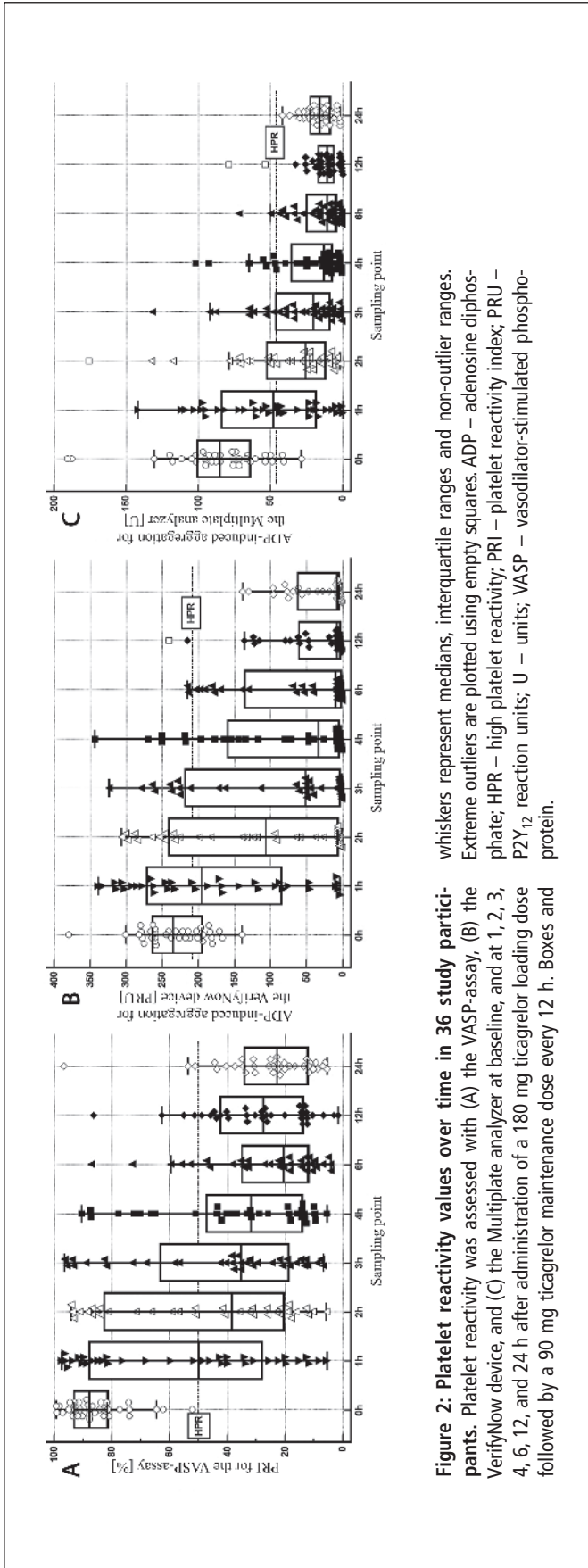


Figure 1: Ticagrelor (A) and AR-C124910XX (B) over time in 36 study participants. Drug concentrations were measured at baseline, and at 1, 2, 3, 4, 6, 12, and 24 h after administration of a 180 mg ticagrelor loading dose

followed by a 90 mg ticagrelor maintenance dose every 12 h. Boxes and whiskers represent medians, interquartile ranges and non-outlier ranges. Extreme outliers are plotted using empty squares.



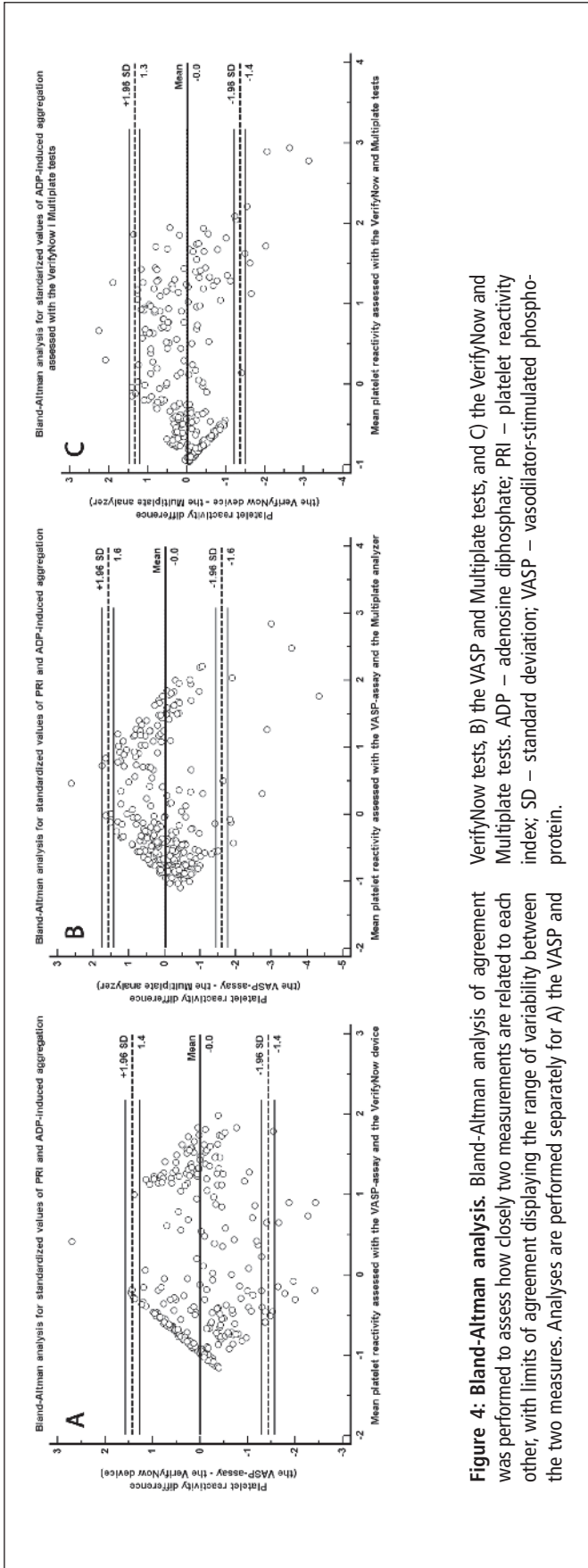


Figure 4: Bland-Altman analysis. Bland-Altman analysis of agreement between VerifyNow tests, B) the VASP and Multiplate tests, and C) the VerifyNow and Multiplate tests. ADP – adenosine diphosphate; PRU – platelet reactivity index; SD – standard deviation; VASP – vasodilator-stimulated phospho-

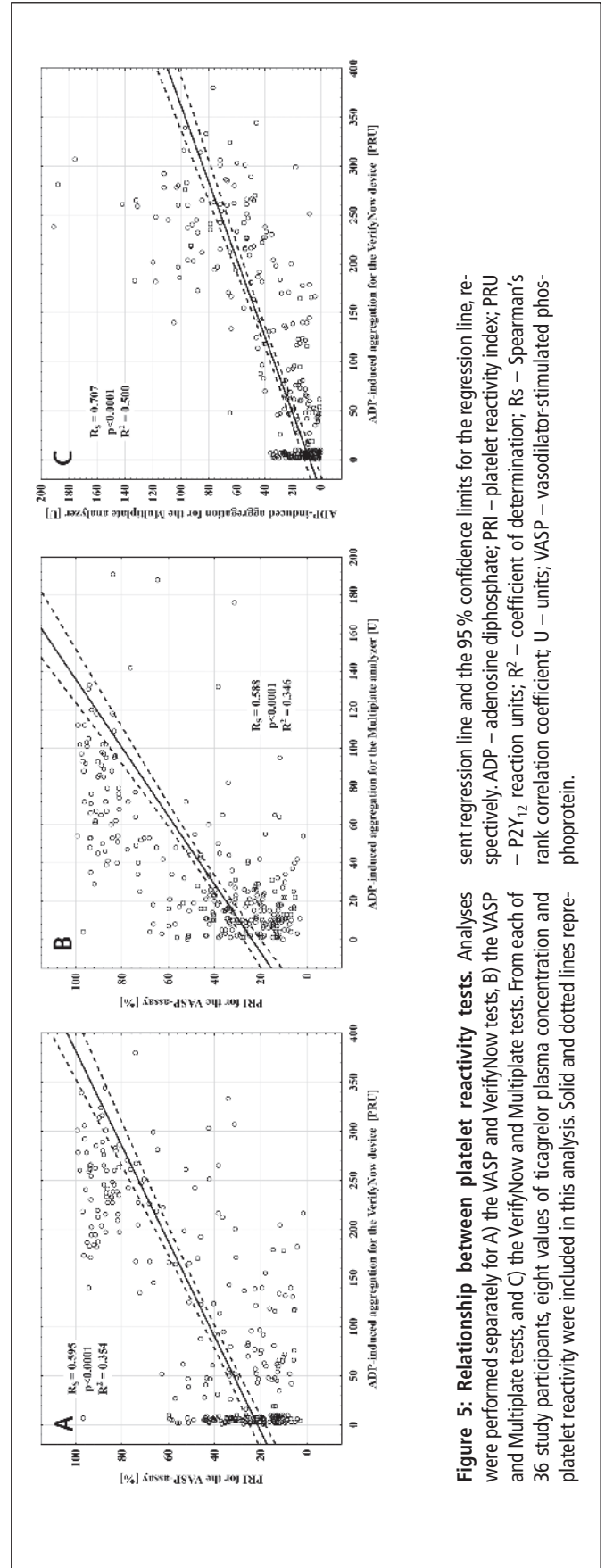


Figure 5: Relationship between platelet reactivity tests. Analyses were performed separately for A) the VASP and VerifyNow tests, B) the VASP and Multiplate tests, and C) the VerifyNow and Multiplate tests. From each of 36 study participants, eight values of ticagrelor plasma concentration and platelet reactivity were included in this analysis. Solid and dotted lines represent regression line and the 95% confidence limits for the regression line, respectively. ADP – adenosine diphosphate; PRU – platelet reactivity index; PRU – P2Y₁₂ reaction units; R² – coefficient of determination; R_s – Spearman's rank correlation coefficient; U – units; VASP – vasodilator-stimulated phosphoprotein.

Rotkreuz, Switzerland). All the above methods were described in detail in our previous publications (12, 13). Platelet reactivity was expressed as platelet reactivity index (PRI), P2Y₁₂ reaction units (PRU), and area under the aggregation curve (AUC) for the VASP-assay, the VerifyNow device and the Multiplate analyzer, respectively. HPR was defined as PRI >50%, AUC >46 units (U), and PRU >208, when assessed with the VASP, Multiplate and VerifyNow tests, respectively (6–8). LPR was identified if AUC was <19 U for the Multiplate analyzer, while due to inconsistencies between expert consensus documents two different cut-off values for LPR were used, for the VASP-assay (<10/<16%) and for the VerifyNow device (<85/<95 PRU) (6–8).

Sample size calculation and statistical analysis

Due to the lack of an appropriate reference study, we decided to perform an internal pilot study on an initial 15 participants to estimate the final sample size for the primary study endpoint. Spearman's correlation coefficient between ticagrelor concentrations and platelet reactivity measured with the VASP-assay was considered a reference for our calculations as this test is regarded as the most accurate tool assessing the effect of thienopyridine or non-thienopyridine inhibitors on the platelet P2Y₁₂ receptor. A value of -0.777 was reached for the first 15 recruited patients. We estimated that a sample size of 35 subjects would have a 95% power to detect a 20% difference in Spearman's correlation coefficient between our reference and values obtained for either the VerifyNow device or the Multiplate analyzer, with a 0.05 two-sided significance level. Values of the lowest detectable differences in

Spearman's correlation coefficient were -0.622 (80% \times -0.777) and -0.932 (120% \times -0.777).

The Shapiro-Wilk test showed a lack of normal distribution for the majority of the analysed quantitative variables, including concentrations of ticagrelor and AR-C124910XX and values of platelet reactivity. Therefore these variables were presented as medians and interquartile ranges (IQR). An extreme outlier was defined as a value that was smaller than the lower quartile minus tripled IQR, or larger than the upper quartile plus tripled IQR. In figures, these values were plotted using a different marker. Depending on the presence or absence of normal distribution, intergroup comparisons were performed with the Student's t-test for independent samples or the Mann-Whitney U test. Unpaired categorical variables were compared using the Chi² test, with Yates' correction if required, or by Fisher's exact test. Correlations were tested with the Spearman's rank correlation test. Coefficients of determination, which are squared correlation coefficients (R²), were also provided as they indicate how well data fit a statistical model. Additionally, intra-class correlation coefficients were calculated. Correlation coefficients were compared using dedicated tests. Agreement between assays to evaluate standardised platelet reactivity was assessed through the Bland-Altman analysis. The κ statistic was used to investigate agreement among the assays in identification of patients with HPR or LPR. A value of two sided $p < 0.05$ was regarded as statistically significant. There were no missing values in our data. The statistical analysis and sample size calculation were carried out using the Statistica 12.0 (StatSoft, Tulsa, OK, USA), MedCalc 15.4 (MedCalc Software, Mariakerke, Belgium) and PQStat 1.6.0.544 (PQStat Software, Poznań/Plewiska, Poland) packages.

Table 2: Agreement between platelet reactivity tests to identify high platelet reactivity as assessed by the κ statistic. Values in the table refer to numbers of measurements, if not stated otherwise. HPR – high platelet reactivity; PRI – platelet reactivity index; PRU – P2Y₁₂ reaction units; U – units.

		VASP-assay		VerifyNow device		Multiplate analyzer	
		HPR absent (PRI \leq 50 %) N=186	HPR present (PRI >50 %) N=102	HPR absent (\leq 208 PRU) N=215	HPR present (>208 PRU) N=73	HPR absent (\leq 46 U) N=205	HPR present (>46 U) N=83
VASP assay	HPR absent (PRI \leq 50 %) N=186			177	9	174	12
	HPR present (PRI >50 %) N=102			38	64	31	71
	κ statistic (95 % CI)			0.623 (0.527; 0.719)		0.655 (0.562; 0.748)	
	Overall agreement between methods			57.7 %		62.3 %	
VerifyNow device	HPR absent (\leq 208 PRU) N=215					196	19
	HPR present (>208 PRU) N=73					9	64
	κ statistic (95 % CI)					0.754 (0.669; 0.840)	
	Overall agreement between methods					69.9 %	

Table 3: Agreement between platelet reactivity tests to identify low platelet reactivity as assessed by the κ statistic. Values in the table refer to numbers of measurements, if not stated otherwise. LPR – low platelet reactivity; PRI – platelet reactivity index; PRU – P2Y₁₂ reaction units; U – units.

		VASP-assay				VerifyNow device				Multiplate analyzer	
		LPR absent (PRI $\geq 10\%$) N=264	LPR present (PRI $< 10\%$) N=24	LPR absent (PRI $\geq 16\%$) N=222	LPR present (PRI $< 16\%$) N=66	LPR absent (≥ 85 PRU) N=129	LPR present (< 85 PRU) N=159	LPR absent (≥ 95 PRU) N=126	LPR present (< 95 PRU) N=162	LPR absent (≥ 19 U) N=155	LPR present (< 19 U) N=133
VASP assay	LPR absent (PRI $\geq 10\%$) N=264					122	142	119	145	146	118
	LPR present (PRI $< 10\%$) N=24					7	17	7	17	9	15
	statistic (95% CI)					0.044 (-0.010; 0.101)		0.040 (-0.016; 0.096)		0.053 (-0.017; 0.122)	
	Overall agreement between methods					10.2%		10.1%		10.6%	
	LPR absent (PRI $\geq 16\%$) N=222					117	105	114	108	138	84
	LPR present (PRI $< 16\%$) N=66					12	54	12	54	17	49
	statistic (95% CI)					0.231 (-0.144; 0.319)		0.220 (-0.134; 0.306)		0.269 (-0.169; 0.370)	
	Overall agreement between methods					31.6%		31.0%		32.7%	
VerifyNow device	LPR absent (≥ 85 PRU) N=129									112	17
	LPR present (< 85 PRU) N=159									43	116
	statistic (95% CI)									0.587 (0.495; 0.678)	
	Overall agreement between methods									65.9%	
	LPR absent (≥ 95 PRU) N=126									109	17
	LPR present (< 95 PRU) N=162									46	116
	statistic (95% CI)									0.567 (0.474; 0.659)	
	Overall agreement between methods									64.8%	

Results

Study population

Between November 2015 and March 2016, 45 AMI patients were enrolled into the study. Baseline characteristics of the study population are presented in ►Table 1. During PCI, nine patients received a GP IIb/IIIa receptor inhibitor and were excluded from the primary analysis. Therefore, pharmacokinetic and pharmacodynamic assessments were performed in 36 patients.

Ticagrelor and AR-C124910XX concentrations and platelet reactivity in the analysed patients

We found a considerable interindividual variability in terms of ticagrelor and AR-C124910XX concentrations and platelet reactivity. Median concentrations of ticagrelor and AR-C124910XX were highest 4 h after the administration of the LD of ticagrelor (►Figure 1). Administration of the 180 mg ticagrelor LD substantially decreased platelet reactivity, which was on average close to the cut-off value for HPR at the 1 h sampling point and reached lowest values between 6 and 24 h after the ticagrelor LD, depending on the method applied (►Figure 2).

Relationship between concentrations of ticagrelor and AR-C124910XX and the results of assessed platelet function tests

We observed high negative correlations between ticagrelor concentrations and platelet reactivity evaluated with all three platelet function tests (►Figure 3), with no significant differences between correlation coefficients. Concordant results were found for AR-C124910XX (Suppl. Figure 1, available online at www.thrombosis-online.com), also with no significant differences between correlation coefficients. Additionally, both ticagrelor and AR-C124910XX concentrations positively correlated with the inhibition of platelet reactivity by ticagrelor, but R^2 values were generally lower than for correlations with platelet reactivity (Suppl. Figures 2 and 3, available online at www.thrombosis-online.com). Consistent with the above results, we failed to find any differences between correlation coefficients determined for the compared methods of platelet reactivity assessment.

Agreement between platelet reactivity tests

The Bland-Altman analysis revealed good agreement between the standardized values of platelet reactivity when measured with all compared platelet function tests (►Figure 4). Importantly, approximately 95% of differences between the standardised values of platelet reactivity in all three comparisons were between the limits of agreement on the Bland-Altman plots.

Platelet reactivity values assessed with all three methods generally correlated well with each other (►Figure 5); however, a significantly higher correlation ($p < 0.02$) was demonstrated between the VerifyNow and Multiplate tests than in other assay combinations. Similar results were obtained for intra-class correlation coefficients between

platelet reactivity when measured with the compared platelet function tests (the VASP-assay and the VerifyNow device: $ICC = 0.737$; $p < 0.000001$; the VASP-assay and the Multiplate analyzer: $ICC = 0.671$; $p < 0.000001$; the VerifyNow device and the Multiplate analyzer: $ICC = 0.762$; $p < 0.000001$), however the difference reached statistical significance only in the comparison between extreme values of intra-class correlation coefficients ($p < 0.02$).

Interestingly, although the κ statistic demonstrated relatively good agreement between all three compared platelet function tests in terms of HPR identification, it was highest between the VerifyNow device and the Multiplate analyzer (►Table 2). On the other hand, for LPR, only results for the VerifyNow and Multiplate test were related to each other (►Table 3).

Comparison of STEMI and NSTEMI patients

Ticagrelor and AR-C124910XX concentrations and values of platelet reactivity over time in STEMI ($n = 22$) versus NSTEMI ($n = 14$) patients are displayed in Suppl. Figures 4 and 5 (available online at www.thrombosis-online.com). Notably, both STEMI and NSTEMI subjects showed considerable inter-individual variability in terms of ticagrelor and AR-C124910XX concentrations and platelet reactivity at all sampling points. Importantly, in STEMI when compared with NSTEMI, we found lower maximum ticagrelor concentration (1167.92 (804.51; 1615.21) ng/ml vs 2342.68 (1299.24; 2511.59) ng/ml; $p < 0.004$) and lower maximum AR-C124910XX concentration (251.97 (175.49; 316.86) ng/ml vs 402.85 (272.55; 638.38) ng/ml; $p < 0.02$), longer time to achieve these two (ticagrelor: 5 (3; 12) h vs 2 (1; 4) h; $p < 0.01$; AR-C124910XX: 6 (4; 12) h vs 3,5 (3; 4) h; $p = 0.082$), as well as delayed maximum antiplatelet effect according to all three methods of platelet function assessment.

Discussion

We believe, this is the first study which demonstrates that all three recommended platelet function tests similarly reflect the *in vivo* plasma concentrations of ticagrelor and its active metabolite. This observation was independently confirmed in two distinct analyses when correlations between drug concentrations and platelet reactivity or platelet inhibition were assessed. Additionally, values of platelet reactivity determined by all three methods generally correlated well with each other; however, a significantly higher correlation was demonstrated between aggregation-based assays, i.e. the VerifyNow and Multiplate tests, than in combinations containing the VASP assay. Importantly, further findings include: i) a relatively good agreement between the recommended platelet function tests in terms of HPR identification, again with the highest measurement concordance being noted between the VerifyNow and Multiplate tests, and ii) a lack of agreement between the compared methods in terms of LPR recognition, except for considerable agreement between the VerifyNow and Multiplate assays. Importantly, our results are largely applicable since ticagrelor is a widely used drug.

Our results suggest that multiple platelet function tests may be no more required in future clinical trials evaluating the antiplatelet effect of ticagrelor. Depending of their local availability, easiness of the use, costs, concomitant therapy with GP IIb/IIIa receptor inhibitors, investigator's preferences, or other factors, one of the tests investigated in our study may be selected for pharmacodynamic studies. We expect that our observations will allow to simplify the design of future studies and lead to resource savings. On the other hand, the rates of on-ticagrelor HPR and particularly LPR determined using currently recommended cut-off values considerably differed between particular platelet function tests in our study and this issue warrants further investigation.

In clopidogrel-treated ACS patients undergoing PCI, HPR and LPR are considered to be risk factors for ischaemic complications, predominantly stent thrombosis, and bleeding events, respectively (5, 14–16). Although some evidence suggests that LPR on ticagrelor is associated with increased risk of bleeding, there is no solid rationale for platelet function testing in patients on ticagrelor therapy (17–19). Apart from superior pharmacokinetic and pharmacodynamic properties of ticagrelor in comparison to clopidogrel, major randomised trials on personalised antiplatelet therapy failed to demonstrate clinical benefits of this approach (20–22).

Previous studies investigating the relationship between clopidogrel active metabolite concentrations and the results of platelet function testing generally favoured the VASP-assay (23–26). Bou-

man et al. found that inhibition of platelet reactivity determined with the VASP-assay ($R^2=0.56$; $p<0.001$) when compared with the VerifyNow device ($R^2=0.48$; $p<0.001$) and light transmission aggregometry (LTA) with 20 $\mu\text{mol/l}$ adenosine diphosphate – ADP ($R^2=0.46$; $p=0.001$ for peak and $R^2=0.47$; $p<0.001$ for late inhibition of platelet reactivity) showed best correlation with peak plasma concentration of the clopidogrel active metabolite (23). Liang et al. also demonstrated superiority of the VASP-assay ($R=-0.58$) over LTA ($R=-0.47$) and the Multiplate analyzer ($R=-0.34$) in terms of correlation with log-transformed concentrations of clopidogrel active metabolite (p for all correlation coefficients <0.01) (24). Additionally, Delavenne et al. observed a higher correlation coefficient between the total area under the clopidogrel active metabolite plasma concentration–time curve from time zero to infinity and inhibition of platelet reactivity for the VASP-assay ($R^2=0.72$; $p=0.008$) than for LTA (10 $\mu\text{mol/l}$ ADP: $R^2=0.49$; $p<0.05$; 5 $\mu\text{mol/l}$ ADP: $R^2=0.61$; $p<0.05$) (25). Importantly, clopidogrel active metabolite is prone to rapid inactivation and irreversibly inhibits platelet function. These facts should be taken into account when interpreting the findings of the above studies.

Our study indicates a delayed and weakened ticagrelor effect in the acute phase of STEMI vs. NSTEMI. However, due to the fact that these findings are derived from an exploratory, post hoc sub-analysis, they warrant in-depth investigation in prospective, powered studies, which are ongoing (26, 27). Importantly, some previous reports indicated that the antiplatelet action of ticagrelor in the STEMI setting may be delayed, but they lacked pharmacokinetic assessment and subjects with other presentations of coronary artery disease as a control group (28, 29). Future studies should particularly address the underlying mechanisms of impaired ticagrelor antiplatelet effect in the STEMI setting and explore its clinical relevance.

Several limitations of our study should be acknowledged. Firstly, only AMI patients admitted to hospital during daytime were enrolled in the study due to difficulties with frequent blood sampling and processing at night. Secondly, patients treated with GP IIb/IIIa receptor inhibitors were excluded from the primary analysis. Thirdly, although we investigated all recommended methods of platelet function assessment (6–8), several other, less validated, assays are also available. Fourthly, despite the fact that HPR and LPR in patients undergoing PCI remain well-documented predictors of ischaemic and bleeding complications, studies linking ticagrelor and AR-C124910XX concentrations with clinical events are lacking. Finally, the study was underpowered for assessment of clinical endpoints, for sub-group analyses and for detection of mild differences between correlation coefficients.

In conclusion, platelet reactivity measurements performed with the VASP-assay, the VerifyNow device and the Multiplate analyzer show comparably strong negative correlations with ticagrelor and AR-C124910XX concentrations. With respect to the recognition of HPR, our study indicates higher measurement concordance between the VerifyNow and Multiplate tests than with other assay combinations, while for LPR, only results of the VerifyNow and Multiplate assays are related to each other.

What is known about this topic?

- Ticagrelor is a direct-acting, reversible platelet P2Y₁₂ receptor inhibitor recommended by both European and American guidelines in patients with acute coronary syndromes. Ticagrelor and its active metabolite (AR-C124910XX) inhibit platelet function faster, more strongly and more consistently than clopidogrel, the former standard of care.
- Multiple platelet function tests are usually simultaneously used in pharmacodynamic studies on P2Y₁₂ receptor inhibitors. According to both European and American experts, there are three recommended platelet function tests, namely the VerifyNow device, the Multiplate analyzer and the VASP assay and none of these three methods is preferred over another.
- So far there are no studies linking measurements of ticagrelor and AR-C124910XX concentrations with comparative evaluation of methods used for platelet reactivity assessment.

What does this paper add?

- This is the first study which demonstrates that all three recommended platelet function tests similarly reflect the *in vivo* plasma concentrations of ticagrelor and its active metabolite.
- Our results suggest that multiple platelet function tests may be no more required in future clinical trials evaluating the antiplatelet effect of ticagrelor.
- Additionally, our study indicates a delayed and weakened ticagrelor effect in the acute phase of STEMI vs. NSTEMI.

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Conflicts of interest

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