

Dual Incorporation of the *in vitro* Data (IC₅₀) and *in vivo* (C_{max}) Data for the Prediction of Area Under the Curve (AUC) for Statins using Regression Models Developed for Either Pravastatin or Simvastatin

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Key words

- pravastatin
- simvastatin
- pharmacokinetics
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- atorvastatin
- rosuvastatin
- AUC
- C_{max}
- IC₅₀

Abstract

Linear regression models utilizing a single time point (C_{max}) has been reported for pravastatin and simvastatin. A new model was developed for the prediction of AUC of statins that utilized the slopes of the above 2 models, with pharmacokinetic (C_{max}) and a pharmacodynamic (IC₅₀ value) components for the statins. The prediction of AUCs for various statins (pravastatin, atorvastatin, simvastatin and rosuvastatin) was carried out using the newly developed dual pharmacoki-

netic and pharmacodynamic model. Generally, the AUC predictions were contained within 0.5 to 2-fold difference of the observed AUC suggesting utility of the new models. The root mean square error predictions were <45% for the 2 models. On the basis of the present work, it is feasible to utilize both pharmacokinetic (C_{max}) and pharmacodynamic (IC₅₀) data for effectively predicting the AUC for statins. Such a new concept as described in the work may have utility in both drug discovery and development stages.

Introduction

Statins are reversible inhibitors of 5-hydroxy-3-methylglutaryl-coenzyme A (HMG CoA) reductase which is a microsomal enzyme responsible for the conversion of HMG CoA to mevalonate [1]. The recent report by Srinivas (2015) has explored the utility of a single time point strategy for predicting area under the curve (AUC) for pravastatin and simvastatin using linear regression models [2]. The applicability of such an approach was also demonstrated to other statins such as atorvastatin, lovastatin and rosuvastatin in a limited manner [1]. In the present work the utility of both pravastatin and simvastatin linear regression models to predict the AUC of other statins was explored. Since statins differ in the *in vitro* potency with regard to the inhibition of HMG co A reductase, it was necessary to use the *in vitro* potency as a surrogate along with the respective *in vivo* C_{max} data of the statin for prediction purposes. This report describes the dual incorporation of pharmacokinetic (C_{max}) and pharmacodynamic (IC₅₀) that has enabled the prediction of AUCs for various statins.

Methods

The slope values of the linear regression models for pravastatin (2.4779) and simvastatin (3.6777) were obtained from the previously published report [2]. The IC₅₀ values for the inhibition of HMG CoA for pravastatin (44.1 nM), simvastatin (11.2 nM), atorvastatin (8.2 nM) and rosuvastatin (5.4 nM) were obtained from the published literature [3,4]. The C_{max} and AUC_{inf} values for the various statins used in the analysis were obtained from the reported pharmacokinetic studies [5–27].

Model development and Predictions using pravastatin and simvastatin slope values

The linear regression model was described by the following relationship that incorporated both C_{max} and IC₅₀ for the prediction purposes. Using pravastatin linear regression slope:

$$AUC(\text{statin} : A) = 2.4779 \times C_{\max}(\text{statin} : A) \times \frac{\sqrt{IC_{50}(\text{pravastatin})}}{\sqrt{IC_{50}(\text{statin} : A)}}$$

Using simvastatin linear regression slope:

$$AUC(\text{statin} : A) = 3.6777 \times C_{\max}(\text{statin} : A) \times \frac{\sqrt{IC_{50}(\text{simvastatin})}}{\sqrt{IC_{50}(\text{statin} : A)}}$$

The predictions of the AUC values with the respective models were carried out on a spread-

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Bibliography

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sheet using Microsoft Excel 2010 (Microsoft Company, Seattle, USA).

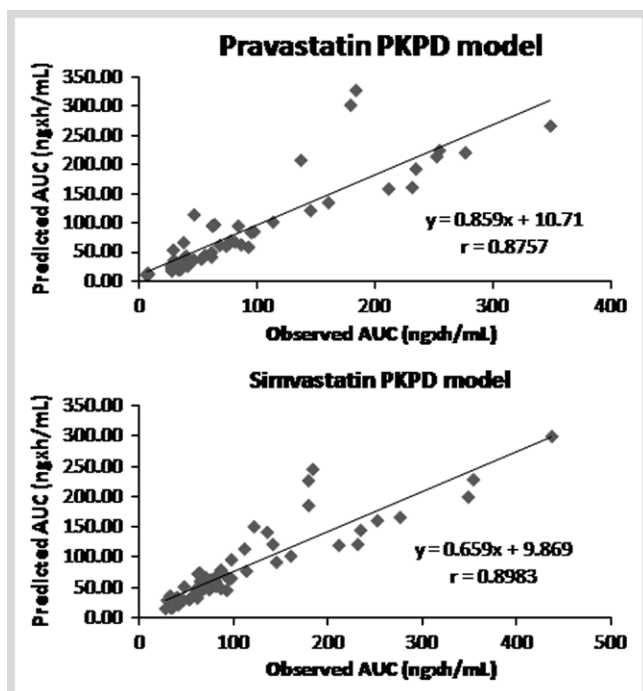


Fig. 1 Plots showing correlation of observed vs. predicted AUC values for various statins using the PKPD model developed for pravastatin and simvastatin.

Fold computation, prediction criteria and statistics

The quotient of observed AUC and predicted AUC value was used to define the fold change of the AUC prediction. The observed vs. predicted AUC values arising from the 2 models (pravastatin or simvastatin) was further evaluated separately by employing a paired t-test (double sided) using the T-test calculator (Graphpad software Inc., California, USA).

The mean absolute error (MAE) was defined as the mean of the observed AUC values minus the predicted AUC values and was computed for both the models:

$$MAE = \sum_{i=1}^N (x_i - y_i)$$

Mean square error (MSE) and root means square error (RMSE) in prediction for both models were performed using Microsoft Excel 2010.

$$MSE = \frac{1}{N} \sum_{i=1}^N (x_i - y_i)^2$$

$$RMSE = \sqrt{\frac{1}{N} \sum_{i=1}^N (x_i - y_i)^2}$$

Results

The AUC predictions rendered by using pravastatin and simvastatin based models are illustrated in Fig. 1, 2, respectively. The

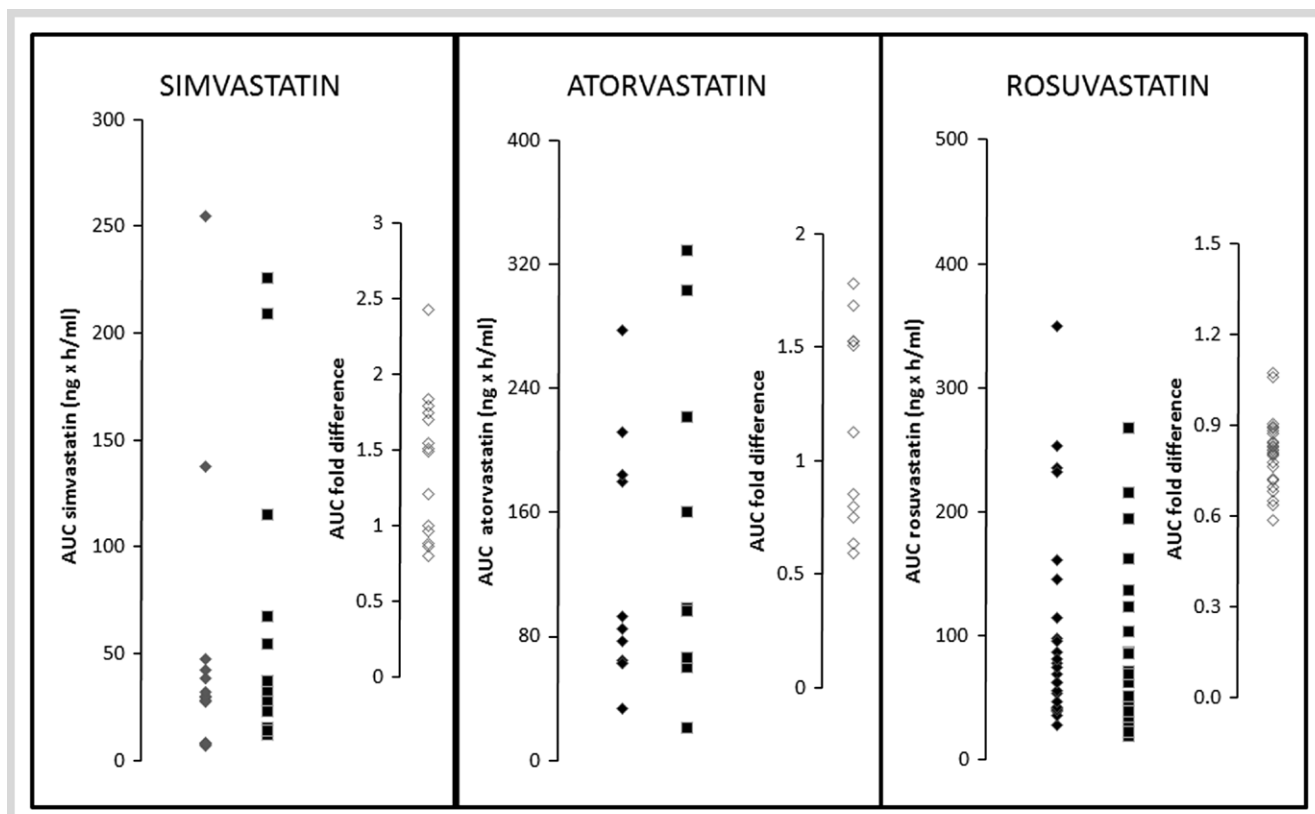


Fig. 2 A plot showing the spread of observed vs. predicted AUC values for simvastatin, atorvastatin and rosuvastatin with the corresponding observed/predicted AUC fold difference. The pravastatin model was used in the analysis [Closed diamonds and closed squares represent the observed and predicted values, respectively; and the open diamonds represent the ratio of the observed AUC/predicted AUC].

predicted values for simvastatin, pravastatin, atorvastatin and rosuvastatin appeared to be comparable based on the visual inspection of the data (► Fig. 1,2). Examination of the AUC fold difference suggested that generally the AUC predictions were contained within 0.5 to 2-fold difference using either of the 2 models (► Fig. 1,2).

The MAE and RMSE expressed as percentage (%) were 2.42 and 42.76, respectively, for the pravastatin based model and the corresponding values were 24.99 and 44.62, respectively, for the simvastatin based model (► Table 1). ► Fig. 3 showed excellent correlations of predicted vs. observed AUC values for various statins regardless of the type of PKPD model employed.

Table 1 Summary statistics for the prediction of AUC values of various statins using the pharmacokinetic-pharmacodynamic model developed for pravastatin and simvastatin.

Model type, (N size)	AUC of statins		Mean absolute error (%)	Root mean Square error (%)
	Observed ³ (ng × h/ml)	Predicted (ng × h/ml)		
Pravastatin ¹ (24)	92.23	90.00	2.23 (2.42)	39.44 (42.76)
Simvastatin ² (36)	108.60	81.46	27.14 (24.99)	48.46 (44.62)

¹ statins included in the analysis were simvastatin, atorvastatin, rosuvastatin

² statins included in the analysis were pravastatin, atorvastatin, rosuvastatin

³ observed values were obtained from the published pharmacokinetic studies (ref: [5–27])

Discussion



In the clinic, a threshold efficacy was achieved for all the statins with differing starting dose sizes. However, the statins that have higher in vitro potency (i.e., atorvastatin and rosuvastatin) tended to show further improvement in the efficacy as the respective dose was increased. Therefore, it was postulated that incorporation of a measure of efficacy (i.e., in vitro potency data for HMG CoA reductase inhibition; IC₅₀ value) along with the respective pharmacokinetic data (i.e., C_{max}) may have utility for the AUC prediction of the statins. Because there was a report of linear regression models developed for both pravastatin and simvastatin, it easily facilitated evaluation of other statins by merely incorporation of C_{max} data of the chosen statin along with the corresponding IC₅₀ value. In the development of the dual PKPD linear regression model, the square root transformation of the IC₅₀ values was found appropriate to give reliable AUC values for the various statins when incorporated in the model. The untransformed “as is” IC₅₀ values generally tended to exhibit higher predictive errors for the various statins and similarly log transformed values appeared to show higher level of deviations (data not shown).

Recently, van de Steeg et al (2013) showed the utility of combining the pharmacokinetic data with the pharmacodynamic data (efficacy data) of the various approved statins in a murine model to enable the translatability of the preclinical model to render

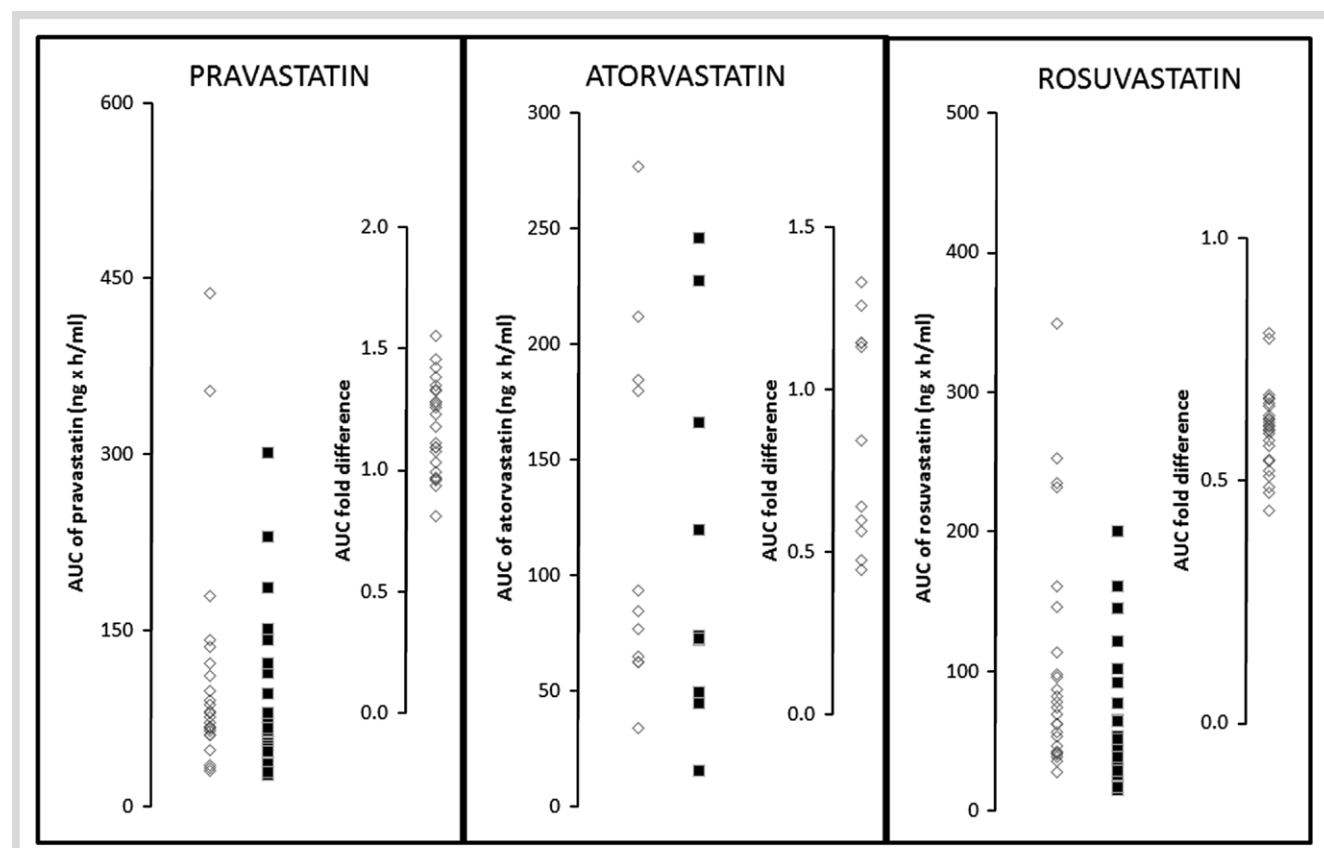


Fig. 3 A plot showing the spread of observed vs predicted AUC values for pravastatin, atorvastatin and rosuvastatin with the corresponding observed/predicted AUC fold difference. The pravastatin model was used in the analysis. The simvastatin model was used in the analysis [Closed diamonds and closed squares represent the observed and predicted values, respectively; and the open diamonds represent the ratio of the observed AUC/predicted AUC].

human predictions [28]. This work showed that incorporation of the effective liver uptake data for the various statins was necessary to improve the translatability of the efficacy in the murine model.

To the best of the author's knowledge, hitherto, the dual incorporation of pharmacokinetic and pharmacodynamic data has not been reported in a linear regression model. Since the 4 statins evaluated in this report showed differences in their IC_{50} value for the inhibition of HMG CoA enzyme, it was thought that incorporation of the IC_{50} value along with C_{max} for each paired statin in relation to the linear regression model developed for pravastatin or simvastatin may render prediction of the AUC of the statin being evaluated. The concept was developed with the view that the intrinsic nature of this class of compounds (i.e., statins) to inhibit HMG CoA enzyme was well established and it was thought that potency of inhibition of the respective statins may be correlated with the appropriate in vivo PK parameter such as C_{max} . The data from the present analysis supported such interesting novel concepts.

Although RMSE values appear to be on the higher side regardless of pravastatin or simvastatin model, it should be noted that 3 other statins with different intrinsic HMG CoA inhibitions and pharmacokinetic profiles were included in the analysis. Therefore, the examples considered for the analysis were not only heterogeneous but also were from different clinical studies including DDI studies. Perhaps, a better control on the RMSE values may be possible if the focus was on a single statin rendering it more homogenous set for prediction purposes. However, it should be noted that the intent of this communication is to suggest a new tool for prediction and from that perspective, the novelty it provides is an important consideration.

One critical aspect that needs to be introspected is what is the rationale in developing such models that incorporate an element of pharmacokinetic (i.e., C_{max}) and pharmacodynamic (i.e., IC_{50}) components in the analysis? Since fast follower approach is commonly followed in the R&D process of new chemical entities (NCE), the development of novel models will be useful for the exposure assessment (AUC) of another new drug within the same chemical class using a different strategy.

Also, because in the drug discovery process, scores of drugs with diversified chemical structures are screened for in vitro efficacy, there is a need for smart and innovative strategy that would enable prediction of exposure from a single time point for making informed decision on the various drug scaffolds or pharmacophores. Typically, primary screens are set to weed out the compounds based on the in vitro target potency and therefore, potency information (if not IC_{50} or K_i values) would be available for the various synthesized compounds. The hit compounds that successfully pass the primary screens may be considered for the same type of analysis reported in the work; however, in this case the developed dual pharmacokinetic-pharmacodynamic model with an anchored reference drug would be based on preclinical rather than clinical data for AUC prediction purposes

Conflict of Interest



The author has stated that he has no conflicts of interest to declare in the contents of this manuscript.

References

- Dietschy JM, Wilson JD. Regulation of Cholesterol Metabolism. *N Engl J Med* 1970; 282: 1128–1138
- Srinivas NR. Limited Sampling Strategy for the Prediction of area under the curve (AUC) of statins: Reliability of a single time point for AUC prediction for pravastatin and simvastatin. *Drug Res (Stuttg)* 2015 [Epub ahead of print]
- Olsson AG, McTaggart F, Raza A. Rosuvastatin: a highly effective new HMG-CoA reductase inhibitor. *Cardiovasc Drug Rev* 2002; 20: 303–328
- McTaggart F, Buckett L, Davidson R et al. Preclinical and clinical pharmacology of rosuvastatin, a new 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor. *Am J Cardiol* 2001; 87 (Suppl): 28B–32B
- Azie NE, Brater DC, Becker PA et al. The interaction of diltiazem with lovastatin and pravastatin. *Clin Pharmacol Ther* 1998; 64: 369–377
- Wu LX, Guo CX, Chen WQ et al. Inhibition of the organic anion-transporting polypeptide 1B1 by quercetin: an in vitro and in vivo assessment. *Br J Clin Pharmacol* 2012; 73: 750–757
- Kyrklund C, Backman JT, Neuvonen M et al. Effect of rifampicin on pravastatin pharmacokinetics in healthy subjects. *Br J Clin Pharmacol* 2004; 57: 181–187
- Triscari J, Swanson BN, Willard DA et al. Steady state serum concentrations of pravastatin and digoxin when given in combination. *Br J Clin Pharmacol* 1993; 36: 263–265
- Pan HY, Triscari J, DeVault AR et al. Pharmacokinetic interaction between propranolol and the HMG-CoA reductase inhibitors pravastatin and lovastatin. *Br J Clin Pharmacol* 1991; 31: 665–670
- Gustavson LE, Schweitzer SM, Koehne-Voss S et al. The effects of multiple doses of fenofibrate on the pharmacokinetics of pravastatin and its 3 α -hydroxy isomeric metabolite. *J Clin Pharmacol* 2005; 45: 947–953
- Pan WJ, Gustavson LE, Achari R et al. Lack of a clinically significant pharmacokinetic interaction between fenofibrate and pravastatin in healthy volunteers. *J Clin Pharmacol* 2000; 40: 316–323
- Krishna R, Garg A, Jin B et al. Assessment of a pharmacokinetic and pharmacodynamic interaction between simvastatin and anacetrapib, a potent cholesteryl ester transfer protein (CETP) inhibitor, in healthy subjects. *Br J Clin Pharmacol* 2009; 67: 520–526
- Bernsdorf A, Giessmann T, Modess C et al. Simvastatin does not influence the intestinal P-glycoprotein and MPR2, and the disposition of talinolol after chronic medication in healthy subjects genotyped for the ABCB1, ABCC2 and SLCO1B1 polymorphisms. *Br J Clin Pharmacol* 2006; 61: 440–450
- O'Brien SG, Meinhardt P, Bond E et al. Effects of imatinib mesylate (STI571, Glivec) on the pharmacokinetics of simvastatin, a cytochrome p450 3A4 substrate, in patients with chronic myeloid leukaemia. *Br J Cancer* 2003; 89: 1855–1859
- Patel CG, Li L, Girgis S et al. Two-way pharmacokinetic interaction studies between saxagliptin and cytochrome P450 substrates or inhibitors: simvastatin, diltiazem extended-release, and ketoconazole. *Clin Pharmacol* 2011; 3: 13–25
- Kosoglou T, Meyer I, Veltri EP et al. Pharmacodynamic interaction between the new selective cholesterol absorption inhibitor ezetimibe and simvastatin. *Br J Clin Pharmacol* 2002; 54: 309–319
- Hsyu PH, Schultz-Smith MD, Lillibridge JH et al. Pharmacokinetic interactions between nelfinavir and 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors atorvastatin and simvastatin. *Antimicrob Agents Chemother* 2001; 45: 3445–3450
- Whitfield LR, Porcari AR, Alvey C et al. Effect of gemfibrozil and fenofibrate on the pharmacokinetics of atorvastatin. *J Clin Pharmacol* 2011; 51: 378–388
- Hulskotte EG, Feng HP, Xuan F et al. Pharmacokinetic evaluation of the interaction between hepatitis C virus protease inhibitor boceprevir and 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors atorvastatin and pravastatin. *Antimicrob Agents Chemother* 2013; 57: 2582–2588 20
- Gajula R, Pilli NR, Ravi VB et al. Simultaneous Determination of Atorvastatin and Aspirin in Human Plasma by LC-MS/MS: Its Pharmacokinetic Application. *Sci Pharm* 2012; 80: 923–940
- Lee JE, van Heeswijk R, Alves K et al. Effect of the hepatitis C virus protease inhibitor telaprevir on the pharmacokinetics of amlodipine and atorvastatin. *Antimicrob Agents Chemother* 2011; 55: 4569–4574
- Pham PA, la Porte CJ, Lee LS et al. Differential effects of tipranavir plus ritonavir on atorvastatin or rosuvastatin pharmacokinetics in healthy volunteers. *Antimicrob Agents Chemother* 2009; 53: 4385–4392
- Jung JA, Lee SY, Kim JR et al. A pharmacokinetic and pharmacodynamic drug interaction between rosuvastatin and valsartan in healthy subjects. *Drug Des Devel Ther* 2015; 9: 745–752

- 24 Allred AJ, Bowen CJ, Park JW *et al.* Eltrombopag increases plasma rosuvastatin exposure in healthy volunteers. *Br J Clin Pharmacol* 2011; 72: 321–329
- 25 Zhang R, Li Y, Jiang X *et al.* Pharmacokinetics and tolerability of multiple-dose rosuvastatin: An open-label, randomized-sequence, three-way crossover trial in healthy Chinese volunteers. *Curr Ther Res Clin Exp* 2009; 70: 392–404
- 26 Keskitalo JE, Kurkinen KJ, Neuvonen M *et al.* No significant effect of ABCB1 haplotypes on the pharmacokinetics of fluvastatin, pravastatin, lovastatin, androsuvastatin. *Br J Clin Pharmacol* 2009; 68: 207–213
- 27 He YJ, Zhang W, Tu JH *et al.* Hepatic nuclear factor 1alpha inhibitor ursodeoxycholic acid influences pharmacokinetics of the organic anion transporting polypeptide 1B1 substrate rosuvastatin and bilirubin. *Drug Metab Dispos* 2008; 36: 1453–1456
- 28 van de Steeg E, Kleemann R, Jansen HT *et al.* Combined analysis of pharmacokinetic and efficacy data of preclinical studies with statins markedly improves translation of drug efficacy to human trials. *J Pharmacol Exp Ther* 2013; 347: 635–644