

# Antibacterial activity of calcium hydroxide and chlorhexidine containing points against *Fusobacterium nucleatum* and *Parvimonas micra*

Andreas Rathke<sup>1</sup>  
Dominik Meisohle<sup>2</sup>  
Jens Bokelmann<sup>3</sup>  
Bernd Haller<sup>1</sup>

## ABSTRACT

**Objective:** Up to 90% of the flora of an infected root canal consists of obligate anaerobic bacteria. However, most studies have centred on microaerophiles. This quantitative assay investigated the antibacterial effects against *Fusobacterium nucleatum* and *Parvimonas micra* of gutta-percha (GP) points containing either calcium hydroxide (Ca(OH)<sub>2</sub>) or chlorhexidine (CHX) compared to those of conventional GP points.

**Methods:** Standardized root canals were prepared in 192 bovine incisors. The canals were sterilized and then inoculated with one of two bacterial species (n=96 teeth per bacterium). After anaerobic incubation at 37°C, the inoculum was removed and the canals were filled with one of the three points. Control teeth were filled with a saline/serum mixture. After 0, 1, 4 and 8 days of anaerobic incubation, the numbers of viable bacteria (CFU/ml) were determined on both the points and the canal wall dentin. Six replicates were carried out for each experiment.

**Results:** None of the points completely eradicated the bacteria in the canal. GP+CHX was significantly more effective than GP+Ca(OH)<sub>2</sub> and GP (P<.05 in each case). The inhibitory effect of GP+Ca(OH)<sub>2</sub> did not significantly differ from that of GP (P>.05). Compared to the controls, the antibacterial effect of the medicated points was reached initially.

**Conclusion:** Within the limitations of the *in vitro* model, the incorporation of commonly used medicaments in GP does not offer a long-lasting antibacterial advantage over non-medicated GP. (Eur J Dent 2012;6:434-439)

**Key words:** Gutta-percha; calcium hydroxide; chlorhexidine diacetate; *Fusobacterium nucleatum*; *Parvimonas micra*; root canal

- <sup>1</sup> Department of Operative Dentistry and Periodontology, University of Ulm, GERMANY
- <sup>2</sup> Division of Oral Microbiology, Department of Operative Dentistry and Periodontology, University of Ulm, GERMANY
- <sup>3</sup> Dentist in Private Practice, Kirchheim unter Teck, GERMANY

■ Corresponding author: Dr. Andreas Rathke  
Department of Operative Dentistry and Periodontology,  
University of Ulm, Albert-Einstein-Allee 11, 89081  
Ulm, GERMANY  
Tel: +49 731 50064155  
Fax: +49 731 50064102  
Email: andreas.rathke@uniklinik-ulm.de

## INTRODUCTION

In microbiologic examinations of teeth with periapical inflammation, evidence of viable bacteria within the infected root canal and adjacent dentinal tubules is frequently found.<sup>1,2</sup> One primary objective in endodontic treatment is the elimination of the bacterial flora in infected teeth. This is usually accomplished by chemo-mechanical preparation of the root canal along with the use of a temporary root-filling between the appointments. Calcium hydroxide (Ca(OH)<sub>2</sub>) is considered to be the endodontic antimicrobial agent of choice for reducing bacteria.<sup>2,3</sup> Chlorhexidine (CHX) seems to be effective in controlling some microorganisms which are not sensitive to Ca(OH)<sub>2</sub>.<sup>4,5</sup> Due to its broad biocidal spectrum, CHX is effective against aerobic and anaerobic bacteria even if it is administered in low concentrations.<sup>6</sup> However, the poor handling properties of the traditional application forms of Ca(OH)<sub>2</sub> and CHX, i.e. liquid suspension, paste or gel, might be one reason why such medicaments often fail to completely disinfect infected root canals.<sup>7</sup> To minimize this problem the use of medicated points that contain the medicaments in a gutta-percha (GP) matrix has recently been recommended.<sup>8</sup> The therapeutic ingredient is released on contact with moisture in the root canal<sup>8</sup> and should therefore be active for a prolonged time.

Dentin and other substances present in the root canal *in vivo* can inhibit the antibacterial efficacies of medicaments.<sup>4,9</sup> For example, Ca(OH)<sub>2</sub> has been reported to completely disinfect *Enterococcus faecalis* in an *in vitro* model.<sup>10</sup> Nevertheless, a review of clinical trials has suggested its limited effectiveness against root canal infections.<sup>3</sup> To date, most of the studies using root canal inoculation models have tested the antibacterial activity of Ca(OH)<sub>2</sub> and/or CHX against aerotolerant or microaerophilic species such as *E. faecalis*.<sup>8,10-13</sup> Little is known about obligate anaerobes, although there are several cultivable species known which may serve as antimicrobial target cells. Furthermore, obligate anaerobic bacteria, such as the Gram-negative *Fusobacterium nucleatum* and the Gram-positive *Parvimonas micra* (formerly *Micromonas micros* or *Peptostreptococcus micros*), account for up to 90% of the endodontic flora in the apical part of infected root canals.<sup>14</sup> These species have been

shown to survive even after the root canal has been permanently filled with GP points and a sealer.<sup>1,14</sup>

The aim of this *in vitro* study was to evaluate the antibacterial effects of GP points containing either additions of Ca(OH)<sub>2</sub> or CHX in comparison to conventional GP points in a modified root canal model inoculated with monocultures of *F. nucleatum* and *P. micra*.

## MATERIALS AND METHODS

### Preparation of Specimens

Bovine, lower central incisors of the same age (n=192) were cut at the level of the cemento-enamel junction using a water-cooled diamond separating disk (Woco 50/Med, Conrad, Clausthal-Zellerfeld, Germany). Standardized root canals measuring 15 mm in length and 0.8 mm in diameter were drilled (Endoseal, Straumann, Basel, Switzerland) in the root dentin, parallel to the axis and the natural root canal of each incisor. The root canals were enlarged with ISO size 80 files (Micro-Mega, Besancon, France) and irrigated with 3 ml of sterile anaerobic 0.9% NaCl. The apex of the roots was shortened with a water-cooled polishing machine (Woco SF 20, Conrad) until the root canal floor was thinned to 0.5 mm. The prepared root specimens were autoclaved at 121°C for 20 minutes and placed in glass vessels containing a freshly prepared mixture of equal amounts of sterile anaerobic 0.9% NaCl solution and bovine serum (B-2771, Sigma, Steinheim, Germany). The cervical part of the root specimens protruded from the medium (Figure 1).

### Inoculation and Medication

The strains *F. nucleatum* American Type Culture Collection (ATCC) 10953 and *P. micra* ATCC 33270 were anaerobically incubated at 37°C in 10 ml of a thioglycolate medium (Oxoid Ltd, Basingstoke, Hampshire, UK) supplemented with 0.5 mg/l vitamin K and 5 mg/l hemin. After incubation overnight, the suspensions were diluted 1:10 with fresh medium and further incubated up to an optical density at 600 nm (OD<sub>600</sub>) of 0.2. With 2 x 10<sup>7</sup> colony-forming units (CFU) per milliliter, this corresponded to an early logarithmic growth phase. The sterile root canals were inoculated with 5 µl of the bacterial suspension corresponding to 1 x 10<sup>5</sup> CFU (n=96 teeth per bacterium). The anaerobic environment was produced by activating BBL GasPak

Plus envelopes (Becton-Dickinson Microbiology Systems, Sparks, USA) in anaerobic jars (GasPak Jars, Becton-Dickinson Microbiology Systems), which were subsequently placed in an incubator at 37°C for 2 days. The inoculum was removed with a sterile micropipette. No additional washing step was performed. The root canals were immediately medicated either with ISO size 80 GP points containing ca 52% Ca(OH)<sub>2</sub> (Calcium Hydroxide Points, Lot number 489808, Roeko Coltène/Whaledent, Langenau, Germany) or ca 5% CHX diacetate (Activ Point, Lot number 399946, Roeko Coltène/Whaledent) or filled with conventional ISO size 80 GP points (Lot number 049808, Roeko Coltène/Whaledent). The production sterility of the points had been verified in earlier studies.<sup>15</sup> Control teeth were filled with the sterile anaerobic NaCl/serum mixture.

### Sampling and Statistical Analysis

After anaerobic incubation at 37°C for 0, 1, 4 and 8 days, each root canal was enlarged with a sterile ISO size 90 file (Micro-Mega). Each file was placed in a sterile tube containing 1.5 ml of sterile anaerobic 0.9% NaCl solution. The capped tube was ultrasonicated for 1 minute and then placed in a centrifuge for 10 minutes at 8000 x g. The sediment of dentinal shavings and bacteria was resuspended in 150 µl NaCl. Following serial dilutions of 10<sup>-1</sup> to 10<sup>-3</sup> in NaCl, 100 µl aliquots were plated onto agar plates and the numbers of CFU/ml were de-

termined as previously described.<sup>16</sup> At the selected time intervals, the points that had been removed were also transferred to sterile tubes containing 1.5 ml NaCl each. Again the numbers of CFU/ml in subsequent dilutions were calculated. Six replicates were carried out for each experiment including the control group. Precautions were taken to ensure asepsis throughout the experiments and to keep the bacteria's exposition to oxygen as low as possible. Preliminary results confirmed that opening the incubation vials frequently and/or for long periods had an unacceptable influence on bacterial growth.<sup>17</sup> Statistical analysis was performed with the nonparametric Kruskal-Wallis test, followed by the Mann-Whitney test. The resulting P values were corrected by using the Holm procedure.

## RESULTS

The antibacterial effect of the three different points against *F. nucleatum* and *P. micra* in the root canal is shown in Figures 2 and 3. For both strains, growth controls showed no significant differences up to 8 days (P>.05). After 1 day, bacterial cell counts in the root canals filled with GP+CHX were significantly lower than in the other groups (P<.05). However, the difference was no longer significant at day 8 for *P. micra* (P>.05). The inhibitory effect of GP+Ca(OH)<sub>2</sub> did not significantly differ from that of GP alone or the growth control at any time interval (P>.05).



Figure 1. Experiments were carried out in glas jars filled with sterile anaerobic NaCl/serum medium and containing the medicated and non-medicated root specimens as well as the bacterial growth controls.

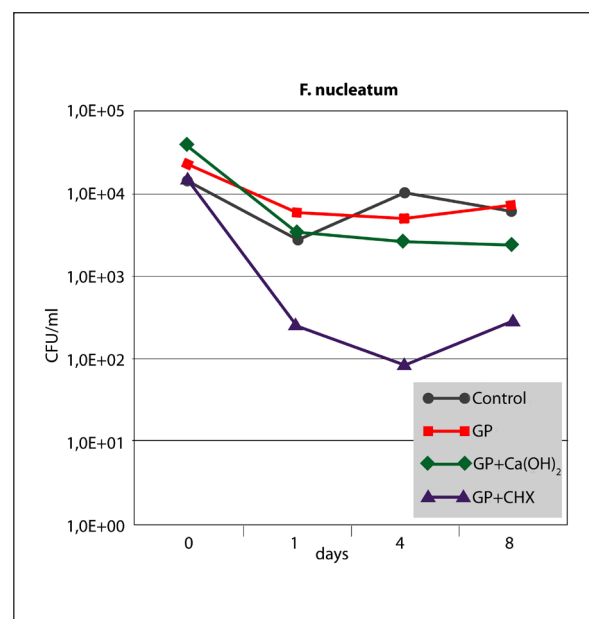


Figure 2. Median CFU/ml counts (n=6) of *F. nucleatum* sampled from inoculated root canals filled with three different points at different time intervals in comparison with the growth control (root canals without points).

When *F. nucleatum* was sampled from the points, GP and GP+Ca(OH)<sub>2</sub> showed constant and high bacterial cell counts over the 8-day period. No significant differences could be observed ( $P < .05$ ). In contrast, GP+CHX showed lower cell counts after one day with bacterial growth increasing on the subsequent days (Figure 4). However, GP+CHX was significantly more effective than GP+Ca(OH)<sub>2</sub> or GP at any time ( $P < .01$ ). Sampling of *P. micra* from the points resulted in similar data (Figure 5). Significant differences were seen between GP+CHX and

GP+Ca(OH)<sub>2</sub> ( $P < .05$ ). No significant differences were found between GP+Ca(OH)<sub>2</sub> and GP ( $P > .05$ ).

## DISCUSSION

In this study, the root canal inoculation test originally described by Haapasalo and Ørstavik was modified to include a quantitative assay and to involve obligate anaerobes as the antimicrobial target cells.<sup>11</sup> Due to the thick dentin layer of bovine incisors, the root canal could be standardized. The smear layer produced during root canal preparation was not removed by irrigation, as this could have distorted the effects of the medicaments.<sup>10</sup> In clinical situations, the antibacterial effect of the medicated GP points may be higher when the root canals are irrigated. In this study, however, the root canals were seeded only for a short period of time. The resulting early phase biofilms are known to be rather susceptible to medicaments compared to a matured biofilm involving complex microbial interactions. When the points tested were removed from the root canals, some of the bacteria were inevitably brushed off on the canal walls. The progressive reaming of the standardized root canal from ISO size 80 to 90 aimed to sample residual bacteria up to dentin depths of 100 µm around the canal lumen. This approach of removing the canal wall dentin with sterile endodontic files and quantitatively determining the viable bacteria by plating the recovered dentin shavings has been described in previous studies.<sup>12, 13</sup> The fact that it was diffi-

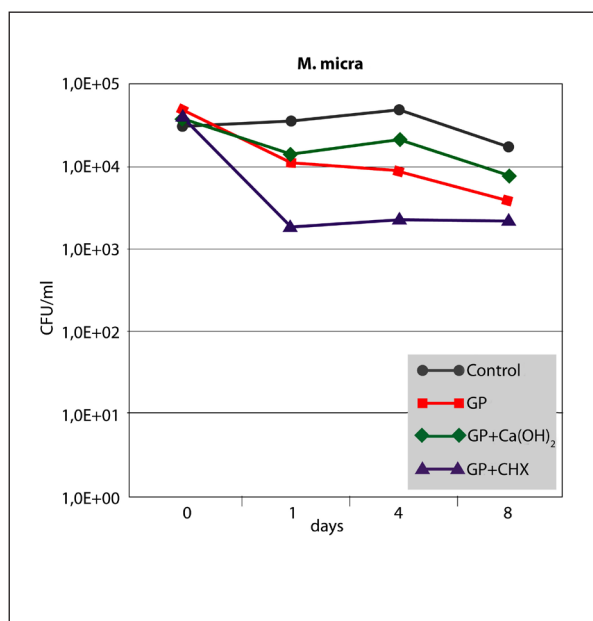


Figure 3. Median CFU/ml counts ( $n=6$ ) of *P. micra* sampled from inoculated root canals filled with three different points at different time intervals in comparison with the growth control (root canals without points).

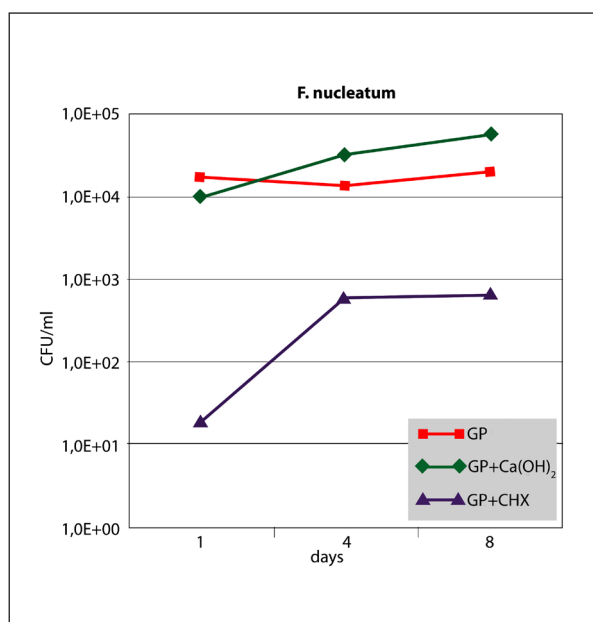


Figure 4. Median CFU/ml counts ( $n=6$ ) of *F. nucleatum* sampled from three different points at different time intervals.

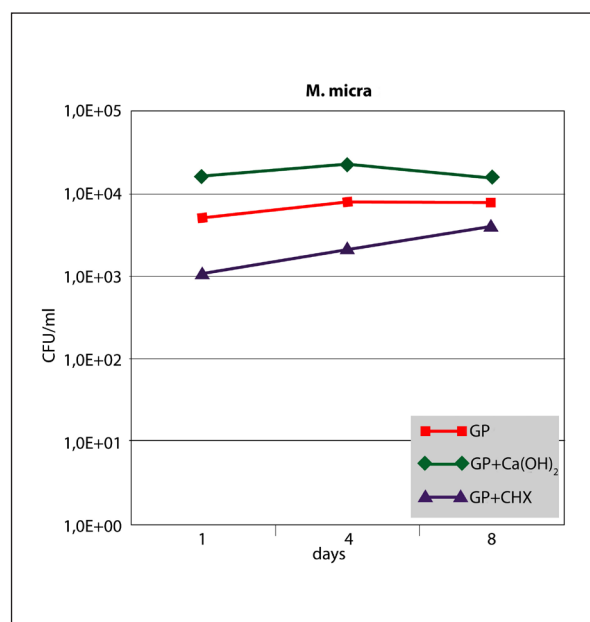


Figure 5. Median CFU/ml counts ( $n=6$ ) of *P. micra* sampled from three different points at different time intervals.

cult to standardize the amount of dentin removed may have contributed to the data variance. However, when the validity was considered, the bacterial samples taken from the root canal and the points showed comparable results. The numbers of CFU/ml from the control teeth were almost constant over the 8-day period with a bacteria load similar to that which is found in a clinically infected root canal.<sup>2,18</sup> The results from the sampling of the growth control at 0, 1, 4 and 8 days after baseline showed that the viable counts of *F. nucleatum* and *P. micra* were not significantly affected by the short-term exposition of the bacteria to oxygen when the incubation vials were opened.<sup>17</sup> Diluted serum was used to simulate the mixture of interstitial liquid and blood filtrate found in clinical situations<sup>5</sup> and to allow any possible influences of plasma proteins on the medicaments (eg, buffering) to be taken into account.<sup>9</sup> This NaCl/serum solution mixture also served as the growth control in earlier studies.<sup>5,16,17</sup>

Although the bacterial strains used are type strains, they were originally isolated from sick humans. According to the information provided by the ATCC product sheet, *F. nucleatum* was isolated from the inflamed gingiva of an adult male and *P. micra* from a patient with purulent pleurisy. Therefore, they should be suitable for early assays for determining the antimicrobial potency of medicated GP points. However, these strains have been used in laboratories over a long period of time and might therefore have lost some of their pathogenic features. Therefore, freshly isolated clinical strains could enhance further tests and might lead to results differing from the present observations.

According to the present results, CHX medicated points were significantly more effective against *F. nucleatum* and *P. micra* than Ca(OH)<sub>2</sub> containing points. This finding supports studies suggesting that CHX has a strong antibacterial effect in concentrations ranging from 0.12% to 5%.<sup>4,12</sup> but contradicts the findings of others reporting similar efficacies for CHX and Ca(OH)<sub>2</sub>.<sup>13,18</sup> The discrepancies in the results may be due to variations in the study design, the concentration and/or the delivery form of the antimicrobial agent. It has been shown that gels with 5% CHX are significantly more effective than CHX containing points.<sup>18</sup> It was surmised that the active ingredients were released from the GP matrix very rapidly. This short-term release may constitute one of the reasons why in the pres-

ent study, the maximum of the antibacterial effect of both CHX and Ca(OH)<sub>2</sub> containing points was reached initially and decreased over the 8-day period.

The minor antibacterial effect of the Ca(OH)<sub>2</sub> containing points in this study coincides with other findings.<sup>19,20</sup> Measurements as a function of time revealed that within 1 day the major amount of Ca(OH)<sub>2</sub> was released from the matrix and the concentration reached a saturation point.<sup>19</sup> A significantly higher pH was only established in the immediate proximity of the Ca(OH)<sub>2</sub> containing points after 1 day and could no longer be detected after 3 days.<sup>20</sup> As a consequence, the alkalinity may not have been high enough to result in the eradication of bacteria. However, the delivery system does not seem to be the only factor that is responsible for the limited effectiveness. Although studies have found Ca(OH)<sub>2</sub> to be more effective when applied as a paste,<sup>18,19</sup> concern has grown about the limited effectiveness of Ca(OH)<sub>2</sub> in general against some species frequently found in clinically infected canals.<sup>3</sup> Despite the fact that there was no significant difference, the Ca(OH)<sub>2</sub> containing points showed a less pronounced effect on *P. micra* than GP in this study. The ZnO of the GP matrix seems to have some antibacterial activity caused by the release of cytotoxic zinc ions.<sup>21</sup> Nevertheless, the fact that chronic apical periodontitis often occurs in conjunction with inadequately filled root canals<sup>2</sup> indicates that GP cannot be expected to lower the bacteria counts significantly in clinically treated root canals.

## CONCLUSION

Under the limitations of the *in vitro* setting, our results suggest that GP loaded with ca 52% Ca(OH)<sub>2</sub> do not possess a long-lasting antibacterial advantage over the non-medicated GP. The incorporation of ca 5% CHX in GP leads to an enhanced antibacterial effect against *F. nucleatum* and *P. micra*, at least in the short-term. However, factors not present in the root canal inoculation model may play an important role in the antimicrobial activity *in vivo*. Particularly, the multispecies communities of biofilms may promote increased resistance levels against antimicrobial agents.

## ACKNOWLEDGEMENTS

The authors deny any financial affiliations related to this study. The authors thank Henry Frehse



(Department of Operative Dentistry and Periodontology, University of Ulm) for the preparation of the root specimens and Prof. Dr. Rainer Muehe (Institute of Biometry, University of Ulm) for the statistical support.

## REFERENCES

- Gomes BP, Pinheiro ET, Gade-Neto C et al. Microbiological examination of infected dental root canals. *Oral Microbiol Immunol* 2004;19:71-76.
- Blome B, Braun A, Sobarzo V, Jepsen S. Molecular identification and quantification of bacteria from endodontic infections using real-time polymerase chain reaction. *Oral Microbiol Immunol* 2008;23:384-390.
- Sathorn C, Parashos P, Messer H. Antibacterial efficacy of calcium hydroxide intracanal dressing: a systematic review and meta-analysis. *Int Endod J* 2007;40:2-10.
- Mohammadi Z, Abbott PV. Antimicrobial substantivity of root canal irrigants and medicament: a review. *Aust Endod J* 2009;35:131-139.
- Podbielski A, Spahr A, Haller B. Additive antimicrobial activity of calcium hydroxide and chlorhexidine on common endodontic bacterial pathogens. *J Endod* 2003;29:340-345.
- Yesilsoy C, Whitaker E, Cleveland D, Phillips E, Trope M. Antimicrobial and toxic effects of established and potential root canal irrigants. *J Endod* 1995;21:513-515.
- Staehele HJ, Thomä C, Müller HP. Comparative in vitro investigation of different methods for temporary root canal filling with aqueous suspensions of calcium hydroxide. *Endod Dent Traumatol* 1997;13:106-112.
- Ebert J, Roggendorf MJ, Frank K, Petschelt A. Antimicrobial activity of various 'active' gutta-percha points against *Enterococcus faecalis* in simulated root canals. *Int Endod J* 2008;41:249-257.
- Portenier I, Waltimo T, Ørstavik D, Haapasalo M. Killing of *Enterococcus faecalis* by MTAD and chlorhexidine digluconate with or without cetrimide in the presence or absence of dentine powder or BSA. *J Endod* 2006;32:138-141.
- Zehnder M, Luder H, Schatzle M, Kerosuo E, Waltimo T. A comparative study on the disinfection potentials of bioactive glass S53P4 and calcium hydroxide in contra-lateral human premolars ex vivo. *Int Endod J* 2006;39:952-958.
- Haapasalo M, Ørstavik D. In vitro infection and disinfection of dentinal tubules. *J Dent Res* 1987;66:1375-1379.
- Schäfer E, Bössmann K. Antimicrobial efficacy of chlorhexidine and two calcium hydroxide formulations against *Enterococcus faecalis*. *J Endod* 2005;31:53-56.
- Lui JN, Sae-Lim V, Song KP, Chen NN. In vitro antimicrobial effect of chlorhexidine-impregnated gutta percha points on *Enterococcus faecalis*. *Int Endod J* 2004;37:105-113.
- Sundqvist G. Taxonomy, ecology, and pathogenicity of the root canal flora. *Oral Surg Oral Med Oral Pathol* 1994;78:522-530.
- Seabra Pereira OL, Siqueira JF Jr. Contamination of gutta-percha and Resilon cones taken directly from the manufacturer. *Clin Oral Investig* 2010;14:327-330.
- Rathke A, Staude R, Muehe R, Haller B. Antibacterial activity of a triclosan-containing resin composite matrix against three common oral bacteria. *J Mater Sci Mater Med* 2010;21:2971-2977.
- Spahr A, Lyngstadaas SP, Boeckh C, Andersson C, Podbielski A, Haller B. Effect of enamel matrix derivate Emdogain on the growth of periodontal pathogens in vitro. *J Clin Periodontol* 2002;29:62-72.
- Barthel CR, Zimmer S, Zilliges S, Schiller R, Göbel UB, Roulet JF. In situ antimicrobial effectiveness of chlorhexidine and calcium hydroxide: gel and paste versus gutta-percha points. *J Endod* 2002;28:427-430.
- Economides N, Koulaouzidou EA, Beltes P, Kortsaris AH. In vitro release of hydroxyl ions from calcium hydroxide gutta-percha points. *J Endod* 1999;25:481-482.
- Schäfer E, Al Behaissi A. pH changes in root dentin after root canal dressing with gutta-percha points containing calcium hydroxide. *J Endod* 2000;26:665-667.
- Silver S. Bacterial resistance to toxic metal ions - a review. *Gene* 1996;179:9-19.