Syntheses of Cyanophycin Segments for Investigations of Cell-Penetration

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Abstract Novel guanidinium-rich oligopeptide derivatives R-[Adp(X)]₈-NH₂ are described, which consist of an octa-aspartic acid backbone with arginylated side chains that are derived from the biopolymer cyanophycin [H-(Adp)n-OH]. The Fmoc-Adp(X,Pbf)-OH building blocks for solid-state peptide synthesis (SSPS) of Adp octamers were prepared from Fmoc-Arg(Pbf)-OH and Fmoc-Asp-OAll. Coupling on PAL resin provided four octamers with and without N-terminal fluorescent groups (FAM) and C-terminal amide groups. Milligram quantities of Adp-octamers were isolated after preparative HPLC purification. The structure of the novel guanidinium-rich oligomers is unique insofar as the side chains of the Asp₈-backbone include both a guanidino and a carboxylic acid group, the influence of which will be tested with the corresponding ester and amide derivatives that were synthesized in parallel. Unusual cell-penetrating properties of the Adp-octamers are expected.

Key words guanidinium-rich oligopeptides, β/α-Asp-Arg-dipeptide building block, biopolymer cyanophycin, solid-state peptide synthesis, cell-penetrating peptides

Arginine- and lysine-rich natural (Tat¹, Penetratin²) and unnatural peptides (oligo-arginines and other guanidinium-rich compounds)³,⁴ are cell-penetrating peptides (CPPs) and can carry a large variety of cargoes into prokaryotic and eukaryotic cells.⁵ A schematic representation is shown in Figure 1A; the number of guanidinium groups is usually between 4 and 14. For a more detailed discussion and additional literature, see the citations in references³⁻⁵ and in the introduction of our recent paper on cell penetration, herbicidal activity, and in vivo toxicity of guanidinium-rich compounds.⁶

Despite all the activity in this field there has been no attention paid by the CPP community, so far, to the biopolymer cyanophycin (Figure 1B), a guanidinium-rich natural product, which was discovered in characteristic granules in blue-green algae by the Italian botanist Antonio Borzi in 1887 and chemically identified by R. D. Simon in 1971.⁷ In recent years, the biopolymer cyanophycin, a temporary microbial nitrogen storage material of cyanobacteria, has been studied most comprehensively by the group of A. Steinbüchel.⁸,⁹ The polymer and its dipeptidic building block can be produced using industrial equipment ‘on any desired scale’.¹⁰ Cyanophycin is a polyaspartic acid arginylated on the carboxylic acid groups of the side-chains, and the building block is a dipeptide with aspartic acid incorporated as a β-amino acid¹¹ (Figure 1B). For simplicity, we use the three-letter code Adp for the cyanophycin building block. Since
peptides with an N-terminal β^-amino acid residue are not cleaved by common aminopeptidases,\textsuperscript{10–12} H-Adp-OH should be quite stable under physiological conditions.

To be able to find out whether cyanophycin segments with a length typical of CPPs (vide supra)\textsuperscript{3–5} have cell penetrating properties we decided to synthesize octamer derivatives (cf. Figure 1 B, with n = 8) by conventional solid-state peptide synthesis (SSPS) using Fmoc chemistry.

For this purpose, the readily available dipeptide H-Adp-OH (Figure 2) looked like a convenient starting material, but this would have required its modification by selectively (!) protecting the guanidino and the carboxylic acid group in the Arg-residue and by putting an Fmoc group on the N-terminus of the Asp residue. Instead, we synthesized suitably protected dipeptide derivatives 1 from the commercially available compounds, Fmoc-Arg(Pbf)-OH (1) and Fmoc-Asp-OAllyl, as outlined in Scheme 1.

The carboxylic acid group of the protected arginine 1 was activated with dicyclohexylcarbodiimide or with thionyl chloride, followed by reactions with t-BuOH, MeOH, or Me$_2$NH to give the protected arginine esters 2a and 2b, and amide 2c, respectively, in yields ranging from ca. 50 to 86%. Removal of the Fmoc group provided the Arg derivatives 3 with free amino groups, to which the Asp moiety was attached by reaction with Fmoc-Asp-OAllyl under peptide-coupling conditions to produce the three Fmoc-Adp(Pbf,X)-OAllyl derivatives 4a-c. De-allylation with phenylsilane/Pd(PPh$_3$)$_4$ led to the building blocks Fmoc-Adp-(O-t-Bu,Pbf)-OH (5), Fmoc-Adp(OMe,Pbf)-OH (6), and Fmoc-Adp(NMe$_2$,Pbf)-OH (7), ready for SSPS (see Scheme 2). Overall yields of up to 40% could be attained for the four steps from Fmoc-Arg(Pbf)-OH (1) to the Adp building blocks 5–7 (for details, see Experimental part). Compound 5 with a t-Bu ester group was actually prepared as precursor to Adp-octamers 8a and 8b with free carboxylic acid groups, formed concomitantly with removal of the peptide from the resin by trifluoroacetic acid (TFA).

As resin for the SSPS we used N-alkylated PAL,\textsuperscript{13} the Fmoc-groups of the growing chains were removed with piperidine in DMF, and the couplings were achieved with HATU/Hünig base (DIPEA) in DMF. The same conditions were employed for attachment of the N-terminal fluorescent FAM label (Scheme 3). Release of the peptide chains from the resin and removal of the Pbf protecting groups was performed with TFA–H$_2$O–TIS, and the products were purified by preparative HPLC. Milligram amounts of the octa-Adp-carboxamides 8–10 (Schemes 2, 3) were synthesized in this way.

We also prepared the octa-Adp-amides with methyl ester 9 and amide groups 10 in the side chains in order to be able to compare the biological activities of Adp-octamers with and without a possible internal neutralization of positively charged guanidinium by negatively charged carboxylate groups (see 8a and b and formulae in Figure 1 B).

The determination of i.v. toxicities and the cell-penetrating properties of octa-Adp derivatives described herein are reported in ref.\textsuperscript{6} and will be described in a separate paper,\textsuperscript{14} respectively.

Protected amino acids and the PAL resin were purchased from Bachem, HATU from Aaptec, 5(6)-carboxyfluorescein (5(6)-FAM) from abcr and all other chemicals were purchased from Sigma Aldrich. All reagents were used as received, solvents were technical grade, and the reactions were run in open flasks fitted with PFTE...
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coated magnetic stir bars at r.t., unless otherwise noted. Peptide couplings were carried out in ISOLUTE® Double fritted filtration column, 15 mL 20 μm PE (reaction vessel, Biotage) at r.t. The building blocks for peptide synthesis were activated in 4 mL screw vial 45 × 14.77 mm (activation vessel, BGB) closed with PFTE lined cap 13-425 (Thermo Scientific) at r.t. Analytical TLC was performed with Merck 60 F254 pre-coated aluminum silica plates and visualized by UV detection (254 nm). Flash column chromatography (FC) was performed using SiliCycle (SilaFlash® P60, 230–400 mesh particle size) silica gel. All fractions collected by FC were analyzed by TLC to identify the different compounds. Melting points were recorded on a Büchi melting point B-540 device.

IR spectra were recorded on a PerkinElmer Spectrum Two FT-IR spectrometer at r.t. using ATR as the sampling technique. NMR spectra were recorded on a Bruker Advance-III 400 MHz spectrometer in the NMR Service at the Laboratory of Organic Chemistry (LOC), ETH Zurich. 1H NMR spectra were recorded relative to the residual solvent peak (CDCl3 δH = 7.26, DMSO-d6 δH = 2.50) and reported as follows: chemical shift (ppm), multiplicity (standard abbreviations; ovlp: overlap), coupling constant (Hz), and integration. 13C NMR spectra were recorded relative to residual solvent peaks (CDCl3 δC = 77.0, DMSO-d6 δC = 39.5). All 1H and 13C signals were assigned via HSQC and HMBC experiments. 19F NMR (D2O) spectra were recorded with 1H decoupling.

The LCMS runs were performed with a Waters Acquity UPLC system equipped with an H-class quaternary solvent manager, an H-class sample manager FTN with sample organizer, a PDA detector, a SQ detector 2, and a 1.7 μm 2.1 × 50 mm BEM C18 UPLC column. Eluent system: H2O and MeCN containing 0.1% HCO2H using a flow of 1 mL/min.

Scheme 2  Deallylation of 4a–c to give the SSPS-building blocks Fmoc-Adp(X,Pbf)-OH 5–7 for assembly to the target octa-Adp derivatives without (8a, 9a, and 10a) and with (8b, 9b, and 10b) a FAM fluorescence label.

Scheme 3  Assemblies of 8 Adp-building blocks by SSPS on PAL resin (Bachem, 200–400 mesh, 0.28 mmol/g) to give the octamers 8–10 with C-terminal amide groups on milligram scale. TIS: Trisopropylsilane.
(Gradient: 0–0.2 min, 5% MeCN; 0.2–1.5 min, 5–80% MeCN; 1.5–2 min, 80–100% MeCN; 2–2.2 min, 100% MeCN; 2.2–2.3 min, 100–5% MeCN; 2.3–3 min, 5% MeCN).

All semi-preparative HPLC runs were performed with a Waters preparative 150 LC system equipped with a 2545 quaternary gradient module, a 2489 UV/visible detector, a Fraction Collector III and 5 μm 20 × 150 mm Reprosil-pure 120 C18 AQ column (Dr. Maisch GmBH, Basel, Switzerland). Eluent system: H2O and MeCN containing 0.1% TFA, using a flow of 10 mL/min (Gradient 1: 0–5 min, 30% MeCN; 5–70, 30–90% MeCN; 70–75 min, 90% MeCN. Gradient 2: 0–5 min, 10% MeCN; 5–45 min, 10–90% MeCN; 45–55 min, 90% MeCN).

All analytical HPLC runs were performed with a Dionex Ultimate 3000 system equipped with a 3000 pump module, a 3000 Autosampler, a 3000 RS Variable Wavelength Detector, and a 3.5 μm 20×150 mm X-Bridge C18 column. Eluent system: H2O and MeCN added with 0.1% TFA, using a flow of 0.1 mL/min (Gradient: 0–1 min, 5% MeCN; 1–16 min, 5–50% MeCN; 16–19 min, 50% MeCN; 19–20 min, 50–85% MeCN; 20–23 min, 95% MeCN).

All self-measured MALDI-TOF were recorded on a Brucker microflex benchtop MALDI-TOF system. High-resolution mass spectrometry (HRMS) was performed on a Bruker maXis UHR-TOF by electrospray ionization (ESI) or a Bruker solariX by matrix-assisted laser desorption/ionization (MALDI) by the Molecular and Biomolecular Analysis Service (MoBiAS) of the LOC at ETH Zurich. MoBiAS also performed elemental analysis.

Preparation of Building Block 5
tert-Butyl N-[(9H-Fluoren-9-yl)methoxy]carbonyl)-N-[(2,2,6,7-pentamethyl-2,3-dihydrobenzofuran-5-yl)sulfonyl]argininate (2a)
In a 100 mL round-bottomed flask, Fmoc-Arg(Pbf)-OH (6.0 g, 9.25 mmol) and DMAP (119.4 mg, 0.977 μmol, 1.1 equiv) were dissolved in DMF (20 mL). Then, t-BuOH (4 mL, 41.82 mmol, 4.52 equiv) was added and the flask was cooled for 5 min on ice. To the cooled mixture, DCC (2.11 g, 10.23 mmol, 1.11 equiv) was added and the resulting solution was stirred for further 5 min on ice. Afterwards, the flask was left with stirring for 3 h 40 min at r.t. The reaction mixture was then poured into 1 M aq HCl (20 mL) and extracted with CH2Cl2 (2 × 20 mL). The organic layers were combined and dried (MgSO4) before being evaporated in vacuo. The crude mixture was purified by FC (EtOAc–CH2Cl2–MeOH, 9:1). All collected fractions were analyzed by TLC, combined and evaporated to give compound 3a (2.08 g, 4.31 mmol, quant.) as a yellowish oil; Rfi = 0.26 (CHCl3–MeOH, 9:1).

1H NMR (400 MHz, CDCl3): δ = 6.32 (t, J = 4.9 Hz, 1 H, Gua NH), 6.15 (s, 2 H, Gua NH), 3.35 (dd, J = 8.3, 4.8 Hz, 1 H, Arg α CH), 3.19 (pent, J = 6.5 Hz, 2 H, Arg CH2), 2.95 (s, 2 H, Pbf CH2), 2.58 (s, 3 H, CH3), 2.52 (s, 3 H, Pbf CH2), 2.09 (s, 3 H, Pbf CH3), 1.83–1.86 (m, 2 H, Arg CH2), 1.70 (s, 2 H, NH2), 1.68–1.49 (m, 2 H, Arg CH2), 1.45 (s, 6 H, Pbf 2 × CH3), 1.45 (s, 9 H, t-CH2).

13C NMR (101 MHz, CDCl3): δ = 171.35 (C=O), 138.58 (Pbf ARC=C), 133.22 (Pbf ARC=C), 132.49 (Pbf ARC=C), 124.70 (Pbf ARC=C), 115.78 (Pbf ARC=C), 86.46 (Pbf C q), 81.75 (t-CH3 C q), 54.52 (Arg α CH), 43.41 (Pbf CH2), 41.09 (Arg CH2), 31.04 (Arg CH2), 28.75 (Pbf CH2), 28.19 (t-CH3 C q), 25.85 (Arg CH2), 19.39 (Pbf CH3), 18.00 (Pbf CH2), 12.61 (Pbf CH3).

LCMS (ESI): m/z = 966.0 (100%) [2 × M + H]+; τi = 1.02 min.


tert-Butyl N-[(2,2,4,6,7-Pentamethyl-2,3-dihydrobenzofuran-5-yl)sulfonyl]argininate (3a)
In a 250 mL round-bottomed flask, compound 2a (3.05 g, 4.33 mmol) was dissolved in piperidine–DMF (50 mL 1:4) and stirred for 80 min. The organic solvent was then evaporated in vacuo and the crude reaction mixture purified 2 times by FC (CH2Cl2–MeOH, 9:1). All collected fractions were analyzed by TLC, combined, and evaporated to give compound 3a (2.08 g, 4.31 mmol, quant.) as a yellowish oil; Rfi = 0.27 (CHCl3–MeOH, 9:1).

1H NMR (400 MHz, CDCl3): δ = 7.75 (dt, J = 7.5, 1.0 Hz, 2 H, Fmoc CH2), 7.60–7.49 (m, 2 H, Fmoc CH2), 7.42–7.34 (m, 2 H, Fmoc CH2), 7.29 (dd, J = 7.4, 4.0, 1.2 Hz, 2 H, Fmoc CH2), 6.14 (s, 1 H, Gua NH), 6.08 (s, 2 H, Gua NH), 5.55 (d, J = 8.1 Hz, 1 H, NH amide), 4.37 (d, J = 7.1 Hz, 2 H, Fmoc CH2), 4.18 (t, J = 7.0 Hz, 2 H, Fmoc CH2 + Arg α CH), 3.35–3.05 (m, 2 H, Arg CH2), 2.91 (s, 2 H, Pbf CH2), 2.58 (s, 3 H, Pbf CH2), 2.51 (s, 3 H, Pbf CH2), 2.07 (s, 3 H, Pbf CH2), 1.87–1.64 (m, 2 H, Arg CH2), 1.64–1.53 (m, 2 H, Arg CH2), 1.45 (s, 9 H, t-CH2), 1.43 (s, 6 H, 2 × Pbf CH3); (EtOAc and traces DCC).

13C NMR (101 MHz, CDCl3): δ = 171.35 (C=O–Cq), 156.23 (Gua Cq), 158.53 (Pbf C q), 133.22 (Pbf C q), 132.49 (Pbf C q), 124.70 (Pbf C q), 115.78 (Pbf C q), 86.46 (Pbf C q), 81.75 (t-CH3 C q), 54.52 (Arg α CH), 43.41 (Pbf CH2), 41.09 (Arg CH2), 31.04 (Arg CH2), 28.75 (Pbf CH2), 28.19 (t-CH3 C q), 25.85 (Arg CH2), 19.39 (Pbf CH3), 18.00 (Pbf CH2), 12.61 (Pbf CH3).

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Synthesis
CH3), 2.54 (3, s, Pbf CH3), 2.50 (3, s, Pbf CH3), 2.09 (3, s, Pbf CH3), 1.91–1.51 (m ovlp, 4 H, 2 × Arg CH2), 1.47 (s, 6 H, 2 × Pbf CH3), 1.45 (s, 9 H, t-C6H5). 13C NMR (101 MHz, CDCl3): δ = 171.47 (C=O amide), 171.01 (C=O allyl ester), 170.77 (C=O t-Bu ester), 161.14 (Pbf amide C=O), 161.31 (Gua amide N=C), 153.81 (Fmoc amide C=O), 149.33 (Fmoc amide C=O), 149.41 (Fmoc amide C=O), 140.95 (Pbf amide C=O), 139.58 (Pbf amide C=O), 131.38 (allyl C=C), 127.88 (Fmoc amide C=O), 127.23 (Fmoc amide C=O), 125.85 (Fmoc amide C=O), 125.29 (Fmoc amide C=O), 120.12 (Fmoc amide C=O), 119.11 (allyl C=C), 118.74 (Pbf amide C=O), 87.58 (Pbf CH3), 83.53 (t-C6H5 CH3), 67.51 (Fmoc CH3), 66.56 (allyl CH2), 51.66 (α CH), 50.80 (α CH), 47.22 (Fmoc CH), 43.06 (Pbf CH), 40.90 (Asp CH2), 37.89 (Arg CH2), 30.93 (Arg CH2), 29.71 (Pbf CH3), 28.05 (t-C6H5 CH3), 24.24 (Arg CH3), 19.38 (Pbf CH3), 17.75 (Pbf CH3), 12.59 (Pbf CH3).

LCMS (ESI): m/z = 862.3 (100%) [M + H]+; tR = 1.73 min. HRMSMS (ESI): m/z [M + H]+ calcd for C35H43N4O7S: 680.389; found: 680.388.

**Synthesis**

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In a 100 mL round-bottomed flask, compound 4b (1.19 g, 1.46 mmol) was dissolved in CH2Cl2 (10 mL) and N2 was bubbled through the solution for 10 min. Then, PhSiH3 (4 mL, 32.45 mmol, 22.25 equiv) and Pd(PPh3)4 were added (66.6 mg, 57.6 μmol, 0.04 equiv). The solution was stirred for 2 h. Then, the mixture was poured into 1 M aq HCl (20 mL) and extracted with CH2Cl2 (3 × 40 mL). The organic layers were combined and evaporated. The crude reaction mixture was separated by FC (CH2Cl2–MeOH, 95:5) and extracted with MeOH (3 × 40 mL). The organic layers were combined and evaporated. Compound 6 was isolated by semi-preparative HPLC with 3 injections using gradient 2 in 51% yield (575 mg, 0.74 mmol) as a colorless powder; mp 118 °C; Rf = 0.29 (MeOH–CH2Cl2, 5:95 + 0.5% TFA).


Preparation of Building Block 7

(9H-Fluoren-9-yl)methyl-(1-dimethylamino)-1-oxo-5-[(2,2,4,6,7-pentamethyl-2,3-dihydrobenzofuran-5-yl)sulfonyl]guanidino]pentan-2-yl)carbamate (2c). Compound 2c (1.19 g, 1.46 mmol) was dissolved in neat SOCl2 (2 mL). The solution was stirred for 30 min. Then, SOCl2 was evaporated using a gentle N2 flow. Afterwards, 2 M HNMe2 in THF (11.5 mL, 22.94 mmol, 10 equiv) was added dropwise by cooling the mixture on ice. The mixture was stirred overnight, 2 M HNMe2 in THF (11.5 mL, 22.94 mmol, 10 equiv) was added dropwise by cooling the mixture on ice. The mixture was stirred overnight. Then, the mixture was poured into 1 M aq HCl (20 mL) and extracted with CH2Cl2 (3 × 40 mL). The organic layers were combined and evaporated. The crude reaction mixture was separated by FC (CH2Cl2–MeOH, 95:5) and extracted with MeOH (3 × 40 mL). The organic layers were combined and evaporated. Compound 6 was isolated by semi-preparative HPLC with 3 injections using gradient 2 in 51% yield (575 mg, 0.74 mmol) as a colorless powder; mp 118 °C; Rf = 0.29 (MeOH–CH2Cl2, 5:95 + 0.5% TFA).


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by TLC, combined, and evaporated to afford compound 2c in 68% yield (1.05 g, 2.29 mmol) as a yellow solid; $R_f = 0.41$ (CH$_2$Cl$_2$-MeOH, 95:5).

1H NMR (400 MHz, CDCl$_3$): $\delta = 7.74$ (d, $J = 7.5$ Hz, 2 H, Fmoc CH$_3$), 7.56 (dd, $J = 7.3$, 3.5 Hz, 2 H, Fmoc CH$_3$), 7.42–7.34 (m, 6 H, 2 × Fmoc CH$_3$), 7.31–7.24 (m, 2 H, Fmoc CH$_3$), 6.21 (s, 1 H, Gua NH), 6.12 (s, 2 H, Gua NH), 6.04 (d, $J = 8.2$ Hz, 1 H, amide NH), 4.72–4.56 (m, 1 H, Fmoc CH$_3$), 4.44–4.26 (m, 2 H, Fmoc CH$_2$), 4.16 (t, $J = 6.9$ Hz, 1 H, Arg $\alpha$ CH$_2$), 3.40–3.08 (m, 2 H, Arg $\beta$ CH$_2$), 3.01 (s, 3 H, NCH$_3$), 2.93 (s, 3 H, NCH$_3$), 2.92 (s, 2 H, Pbf CH$_2$), 2.58 (s, 3 H, Pbf CH$_3$), 2.51 (s, 3 H, Pbf CH$_3$), 2.08 (s, 3 H, Pbf CH$_3$), 1.81–1.66 (m, 2 H, Arg CH$_2$), 1.61 (dd, $J = 13.8$, 7.1 Hz, 2 H, Arg CH$_2$), 1.43 (s, 6 H, 2 × Pbf CH$_3$).

HRMS (ESI): $m/z$ calcd for C$_{63}$H$_{94}$N$_{12}$O$_{14}$S: 1327.655; found: 1327.654.

2-Amino-N,N-dimethyl-5-[3-[(2,2,4,6,7-pentamethyl-2,3-dihydrobenzo[5-5]thiazol-2-yl)sulfonyl]guanidino]pentamidine (3c)

In a 100 mL round-bottomed flask, compound 2c (898 mg, 1.46 mmol) was dissolved in piperidine–DMF (1:4, 10 mL). The solution was stirred for 1 h. Then, 1 M aq HCl (30 mL) was added, and the mixture was transferred to the 100 mL flask and the resulting solution was stirred overnight. Finally, the reaction mixture was poured into 1 M aq HCl (20 mL) and extracted with CH$_2$Cl$_2$ (3 × 40 mL). The organic layers were combined, dried (MgSO$_4$), and evaporated to afford compound 3c (664 mg, 1.46 mmol) dissolved in DMF (8.8 mL). In another small pear-shaped flask, Fmoc-Asp-OAll (607.8 mg, 1.54 mmol, 1.05 equiv) was dissolved in 0.4 M HATU in DMF (3.8 mL, 1.54 mmol, 1.05 equiv). Then, DIPEA (524 µL, 3.07 mmol, 2.1 equiv) was added, and the flask was shaken for 2 min. Afterwards, the content of the pear-shaped flask was transferred to the 100 mL flask and the resulting solution was stirred overnight. Finally, the reaction mixture was poured into 1 M aq HCl (20 mL) and extracted with CH$_2$Cl$_2$ (3 × 40 mL). The organic layers were combined, dried (MgSO$_4$), and evaporated in vacuo. The remaining DMF was removed using high vacuum (10$^{-2}$ mbar). The crude reaction mixture was then purified by FC (CH$_2$Cl$_2$–MeOH, gradient 100:0 to 95:5). The fractions containing the product (TLC analysis) were combined and evaporated in vacuo. Further purification via FC (CH$_2$Cl$_2$–MeOH, 96:4) and TLC analysis of the collected fractions gave, after evaporation, compound 4c in 77% yield (938 mg, 1.13 mmol) as a colorless solid; $R_f = 0.30$ (MeOH–CH$_2$Cl$_2$, 9:1).

1H NMR (400 MHz, CDCl$_3$): $\delta = 7.76$ (d, $J = 7.6$ Hz, 2 H, Fmoc CH$_3$), 7.60 (t, $J = 6.2$ Hz, 2 H, Fmoc CH$_3$), 7.40 (t, $J = 7.5$ Hz, 2 H, Fmoc CH$_3$), 7.30 (t, $J = 7.5$ Hz, 2 H, Fmoc CH$_3$), 7.22 (d, $J = 7.0$ Hz, 1 H, amide NH), 6.58 (s, 3 H, Gua NH), 5.94 (d, $J = 7.3$ Hz, 1 H, amide NH), 5.85 (d, $J = 10.8$, 5.2 Hz, 1 H, allyl C=CH$_2$), 5.29 (d, $J = 17.1$ Hz, 1 H, allyl C=CH$_2$), 5.23 (d, $J = 10.5$ Hz, 1 H, allyl C=CH$_2$), 4.80 (t, $J = 7.9$ Hz, 1 H, $\alpha$ CH), 4.73–4.65 (m, 1 H, $\alpha$ CH), 4.65–4.53 (m, 2 H, $\beta$ CH$_2$), 4.48–4.16 (m, ovlp, 3 H, Fmoc CH + Fmoc CH$_2$), 3.36 (d, $J = 154.1$ Hz, 2 H, Arg CH$_2$), 3.02 (s, 3 H, NCH$_3$), 2.96 (s, 3 H, NCH$_3$), 2.95 (s, 2 H, Pbf CH$_3$), 2.14–2.80 (m, 2 H, Asp CH$_2$), 2.55 (s, 3 H, Pbf CH$_3$), 2.51 (s, 3 H, Pbf CH$_3$), 2.00 (s, 3 H, Pbf CH$_3$), 1.64 (dd, ovlp, $J = 24.2$, 18.0, 11.3 Hz, 4 H, 2 × Arg CH$_2$), 1.46 (s, 6 H, 2 × Pbf CH$_3$).

HRMS (ESI): $m/z = 833.5$ (100% $[M + H]^+$; $t_f = 1.64$ min.

HRMS (ESI): $m/z$ calcd for C$_{65}$H$_{96}$N$_{20}$O$_{21}$S: 831.3746; found: 831.3731.


In a 100 mL round-bottomed flask under N$_2$, compound 4c (882 mg, 1.06 mmol) was dissolved in CH$_2$Cl$_2$ (10 mL) and N$_2$ was bubbled through the solution for 10 min. Then, Pd(PPh$_3$)$_4$ was added (126.9 mg, 109.8 µmol, 0.1 equiv) and again N$_2$ was bubbled through the mixture for 2 min. Afterwards, PhSiH$_3$ (2.8 mL) was added and the reaction mixture was stirred for 2 h. Finally, the mixture was poured into 1 M aq HCl (20 mL) and extracted with CH$_2$Cl$_2$ (3 × 40 mL). The organic layers were combined and dried (MgSO$_4$) before being evaporated in vacuo. Compound 7 was isolated via semi-preparative HPLC with 3 injections and gradient 2 in 61% yield (510 mg, 0.64 mmol) as a colorless powder; mp 145 °C.
IR (neat): 3330 (w), 2971 (w), 2934 (w), 1719 (m), 1667 (m), 1627 (s), 1576 (m), 1549 (m), 1450 (m) 1408 (m), 1372 (w), 1322 (w), 1292 (m), 1254 (m), 1202 (m), 1165 (m), 1135 (s), 1090 (s), 1059 (m), 996 (m), 850 (m), 782 (m), 760 (s), 741 (s), 700 (m), 641 (s), 620 (m) cm⁻¹.

1H NMR (400 MHz, DMSO-d₆): δ = 8.13 (d, J = 8.2 Hz, 1 H, amide NH), 7.88 (d, J = 7.6 Hz, 2 H, Fmoc CH₃), 7.70 (d, J = 7.4 Hz, 2 H, Fmoc CH₂), 7.50 (d, J = 8.4 Hz, 1 H, amide NH), 7.41 (td, J = 7.5, 1.1 Hz, 2 H, Fmoc CH₃), 7.32 (dt, J = 7.5, 1.5 Hz, 2 H, Fmoc CH₃), 6.64 (s, 1 H, Gua NH), 6.40 (s, 1 H, Gua NH), 4.65 (q, J = 7.9 Hz, 1 H, Fmoc CH), 4.34 (td, J = 8.1, 5.5 Hz, 1 H, α CH), 4.30–4.15 (m ovlp, 3 H, Fmoc CH₂) + (DMSO signal), 3.86–2.97 (m, 2 H, Arg CH₂), 2.95 (s, 3 H, NCH₃), 2.93 (s, 2 H, Pbf CH₂), 2.80 (s, 3 H, NCH₃), 2.56 (dd, J = 17.0, 6.6 Hz, 2 H, Asp CH₂), 2.46 (s, 3 H, Pbf CH₃), 2.41 (s, 3 H, Pbf CH₃), 1.99 (s, 3 H, Pbf CH₃), 1.66–1.50 (m, 2 H, Arg CH₂), 1.39 (s, 6 H, 2 × Pbf CH₃), 1.48–1.25 (m, 2 H, Arg CH₂).

13C NMR (101 MHz, DMSO-d₆): δ = 172.99 (δ C=O amide), 170.88 (δ C=O NMe₂ amide), 168.73 (CO₂H), 157.42 (Pbf CH₂), 156.03 (Fmoc C=O), 152.07 (Fmoc CH₃), 143.78 (Fmoc μC=O), 144.75 (Fmoc μC=O), 140.69 (Fmoc μC=O), 137.25 (Pbf μC=O), 134.14 (Pbf μC=O), 131.42 (Pbf μC=O), 127.62 (Fmoc μC=O), 127.06 (Fmoc μC=O), 125.24 (Fmoc μC=O), 124.31 (Pbf μC=O), 120.09 (Fmoc μC=O), 116.24 (Pbf μC=O), 86.28 (Pbf CH₃), 65.72 (Fmoc CH₂), 50.67 (α CH), 48.04 (Fmoc CH₂), 46.58 (α CH), 42.44 (Pbf CH₂), 39.73 (Arg CH₂ ovlp with DMSO signal), 36.75 (Asp CH₂), 36.50 (NCH₃), 35.16 (NCH₃), 28.95 (Arg CH₂), 28.28 (Pbf CH₃), 25.09 (Arg CH₂), 18.94 (Pbf CH₂), 17.58 (Pbf CH₃), 12.26 (Pbf CH₃).

LCMS (ESI): m/z = 791.9 (100%) [M + H]+; tᵣ = 1.61 min.

HRMS (ESI): [M + H]+ calcd for C₉₈H₃₂N₄₀O₅₀ (hygroscopic!): C, 39.79; H, 4.66; F, 0.43; N, 15.33. Found: 2298.7030.

This procedure describes the cleavage from the resin for peptides 8–10. After the last coupling, the Fmoc group was removed by using the same procedure as described in GP1. The resin was then washed 5 × 1 min with DMF (5 mL each) and then 5 × 1 min with CH₂Cl₂ (5 mL each). Finally, the resin was dried by suction for approximately 25 min.

The dry resin was transferred to a tared 100 mL round-bottomed flask and weighed. Per g of dried resin, 50 mL of cleavage cocktail was added [R-{[Adp(OH)]₈-NH₂ (8): TFA–TIS–H₂O 95:2.5:2.5; R-{[Adp(OCH₃)]₈-NH₂ (9), and R-{[Adp(NMe₂)]₈-NH₂ (10): TFA–TIS–MeOH 95:2.5:2.5}] and the reaction mixture was stirred at r.t. under N₂ for 4 h.

Afterwards, the resin was filtered off using a fritted glass filter and washed with neat TFA. The TFA was evaporated with N₂ flow until some material started to precipitate. Then, ice cold Et₂O was added and the resulting suspension was filtered on Celite and rinsed with ice cold Et₂O to remove cleaved protecting groups. To solubilize the peptide material, the Celite was resuspended 3 × in H₂O–MeCN + 0.1% TFA (40 mL each, 1:1) and filtered. The filtrate was frozen, lyophilized, and stored at 4 °C before purification via semi-preparative HPLC (gradient 1). All HPLC fractions were analyzed by analytical HPLC, combined according to purity (all samples considered >95% pure), and lyophilized to yield the TFA salt form of the peptides. Peptides 8a, 9a, and 10a were obtained as colorless powders (5–15 mg each; hygroscopic). The TFA content of the salt was determined by elemental analysis of 8a, according to which 9 TFA molecules were associated with the peptide. We assumed that this was also the case for 9a and 10a. For a typical desalting procedure, vide infra.

**Preparation of the Octamers 8–10**

**Solid-State Peptide Synthesis; General Procedure (GP1)**

All cyanophycin octamer derivatives were synthesized using the following protocol on a 0.03 mmol scale (building block 5 for 8a,b; 6 for 9a,b; and 7 for 10a,b).

In a 15 mL reaction vessel equipped with a valve and attached to a suction system, ca. 107 mg of dry PAL resin (0.28 mmol/g) was shaken in DMF (5 mL) for 30 min. Then, the resin was treated with piperidine–DMF (1:4, 5 mL each) for 8 min) with piperidine–DMF (1:4, 5 mL each). The resin could then be stored for a few days at 4 °C in a sealed reaction vessel before cleavage (to give 8a, 9a, and 10a) (see GP2, vide infra) or FAM coupling (to give 8b, 9b, and 10b) (see FAM coupling procedure, vide infra).
ed and the mixture was shaken for 10 min. The content of the activation vessel was then transferred to the reaction vessel and shaken overnight (16 h). The resin was then washed 5 x 1 min with DMF (5 mL each) and treated with piperidine–DMF (1:4, 5 mL) for 30 min. After this step, resin-bound 8b, 9b, and 10b were submitted to cleavage following GP2. The TFA salts of the FAM derivatives 8b, 9b, and 10b were obtained as yellow powders (2–3 mg each; hygroscopic).

8b
HPLC: t_R = 8.4 min.
MS (MALDI): m/z = 2548.919 (100%) [M + H]^+.

9b
HPLC: t_R = 9.8 min.
MS (MALDI): m/z = 2657.685 (100%) [M + H]^+.

10b
HPLC: t_R1 = 9.1 min, t_R2 = 9.2 min.
MS (MALDI): m/z = 2761.782 (100%) [M + H]^+.

Note: For FAM-[Adp(OH)]_8-NH_2 (8b) and FAM-[Adp(OMe)]_8-NH_2 (9b), one constitutional isomer of the FAM labeled peptide could be isolated by semi-preparative HPLC (5-FAM or 6-FAM). In the case of FAM-[Adp(NMe_2)]_8-NH_2 (10b), only a mixture of the 2 isomers (5/6-FAM) could be isolated in pure form (see Figure 3).

Desalting the Octapeptides; Typical Procedure
Desalting of 8a
In a 50 mL falcon tube, a solution of 8a in H_2O (5 mL, double deionized) was incubated at r.t. for 10 min with Amberlyst A26 (HO form, 1.5 g). The mixture was then transferred to a fritted glass filter and washed 5 times with H_2O (each 20 mL, double deionized). The filtrate was collected, frozen, and lyophilized in order to give desalted 8a. For control of fluorine content a ¹⁹F NMR spectrum was recorded; there was only a tiny little signal from TFA (δ = –75.60).

References